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1	Combined ultrasonication and thermal pre-treatment of sewage sludge for increasing
2	methane production
3	
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29 Abstract

30 This paper focuses on the combination of ultrasonic and thermal treatment of sewage sludge (SS). 31 The combination involved ultrasonicating a fraction of the sludge and thermal treatment at various temperatures and this resulted in solubilization of proteins and carbohydrates, and so 32 33 contributing to increased COD solubilization. During the treatment, SCOD, soluble proteins and 34 carbohydrates increased from 760 mg/L to 10,200 mg/L, 110 mg/L to 2,900 mg/L and 60 mg/L to 35 630 mg/L, respectively. It was found ultrasonication of only a fraction of the sludge (>20%) 36 followed by thermal treatment led to significant improvement compared to thermal and ULS 37 treatments applied on their own. At 65°C, the kinetic of solubilization was improved and the 38 hyper-thermophilic treatment time could be reduced to a few hours when ultrasonication was used first. A linear correlation ($R^2 = 95\%$) was found between the SCOD obtained after ultrasonication 39 40 pre-treatment and anaerobic biodegradability. The combined treatment resulted in 20% increase in 41 biogas production during the anaerobic digestion of the pre-treated sludge.

42

43 Keywords: Waste Activated Sludge (WAS); ultrasonication; thermal pre-treatment; anaerobic
44 biodegradability.

45

46 Introduction

48 Large amount of surplus biological sludge is generated during activated sludge process. Cost for 49 sludge treatment and disposal may take as high as 50% of total cost for a wastewater treatment plant.^[1] Anaerobic digestion is commonly accepted as an ideal method to stabilize sludge for safe 50 disposal and utilization.^[2] It has the advantages of low biomass yield, high stabilization degree as 51 well as production of methane gas.^[3] It is known that the digestible organic fraction in Waste 52 53 Activated Sludge (WAS) is only about 30-45% (w/w) of biomass in conventional anaerobic 54 treatment, while the methane production can improve markedly by disintegrating the WAS cells to release the intracellular organics using chemical or mechanical disruption methods.^[4] 55 56

57 WAS mainly consists of intact microorganisms and their secretions forming particles larger than 58 0.1 µm that cannot be directly assimilated by the microorganisms. Hydrolysis of the cells must 59 first take place before the soluble materials released can be converted to methane gas in the 60 anaerobic digester. Cell lysis of the microorganisms limits the rate of hydrolysis which further limits the rate of the whole anaerobic process. ^[5] Furthermore, during the activated sludge 61 62 process, bacterial cells form flocs which structure is enhanced by extracellular polymeric 63 substances (EPS). This complex structure protects microorganisms from being degraded and 64 makes the cell lysis even harder.

Pre-treatment of WAS has been proven to disrupt sludge structures, causing solubilisation of organics and accelerate subsequent anaerobic digestion. ^[6-9] One of the pre-treatment method is an anaerobic or aerobic biological method that requires either thermophilic (around 55°C) or hyperthermophilic (between 60 and 70°C) conditions (Table 1) which typically result in an increase in hydrolysis activity, increase of biodegradable COD and pathogen destruction. ^[4-10-11] The increase in hydrolytic activity was demonstrated by Hasegawa et al. ^[12] who reported 40% VSS

solubilization due to the pre-treatment. Production of biogas after anaerobic digestion of the
microaerobically-pretreated sludge was increased by 1.5X when compared with the sludge
without pre-treatment. Destruction of 75% organic solids from excess waste activated sludge was
obtained at full scale, by combining a conventional municipal activated sludge process with a
thermophilic aerobic sludge digester (65°C, HRT of 2.8 days).^[13] However, depending on the type
of sludge (primary, secondary or mixture of both) the residence of this type of treatment is
generally 2 days or longer.

78 Another relatively new pre-treatment of WAS is ultrasonication. Huge hydro-mechanical shear 79 force generated by cavitation bubbles during ULS is believed to be the predominant effect for sludge disintegration.^[7-14] In contrast with thermal methods, ULS is, comparatively, a very rapid 80 81 method that causes solubilization of both extracellular and intracellular substances leading to an increase in soluble microbial products.^[15] Microorganisms in WAS degrade the organic matter by 82 83 producing hydrolytic enzymes that are released into the media. Therefore, a physical treatment 84 such as ULS should be useful to disrupt the flocs, release the enzymes and at the same time 85 improve the thermal pre-treatment, but information on combination of these two pre-treatment methods is still scarce in the literature. Since ultrasonication is an energy-intensive process, its 86 major disadvantage is its high energy consumption.^[16] Therefore, the ultrasonication of a fraction 87 88 of WAS would be an interesting option to study and the objective of this paper was to investigate 89 how combination of ULS and thermal pre-treatments can enhance methane production. The 90 specific objectives were to determine the optimum temperature and duration of thermal pre-91 treatment of WAS, and to determine how ultrasonication could improve solubilization of COD, 92 proteins and carbohydrates when it is applied before thermal treatment. Another objective was to

93	investigate the	solubilization	of WAS	when on	ly a fraction	n of it wa	s ultrasonicated	l and to
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94 determine the impact on solids destruction and biogas production.

95

96 [Table 1]

97

98 Materials and methods

99

100 Sludge samples

101

102 Mixture of primary sludge and thickened waste activated sludge (ratio around 1:1 based on dry

103 solids) were collected from Ulu Pandan municipal wastewater reclamation plant (Singapore).

104 Properties of the sludge used in this study are listed in Table 2.

105 **[Table 2]**

106

107 Analytical methods

- 109 The measurement of pH (Jenway) was accurate to within ±0.02 units. The Total Solids (TS),
- 110 Volatile Solids ^[17], Total Suspended Solids (TSS), Volatile Suspended Solids (VSS), Soluble
- 111 Chemical Oxygen Demand (SCOD) and Total Chemical Oxygen Demand (TCOD) were
- 112 measured in triplicate as described in Standard Methods.^[18] Their coefficient of variation ^[19] for
- 113 ten identical samples was 2.7%, 3.8%, 2.8%, 4.8%, 1.9 and 1.6%, respectively.

114	Proteins concentration was determined in triplicate by Lowry's method ^[20] using bovine serum
115	albumin (Sigma-Aldrich) as standard and a UV/VIS scanning spectrophotometer (Shimadzu, UV-
116	1800) against the blank at a wavelength of 750 nm. The coefficient of variance was within 2.8%
117	for ten identical samples. As the precise chemical formula of the proteins detected was not
118	determined, the percentage of soluble COD represented by protein had to be estimated by
119	assuming a stoichiometric conversion factor of 1.5 which is derived from the typical formula of
120	proteins (C ₁₆ H ₂₄ O ₅ N ₄) presented in Rittmann and McCarty. ^[21] Carbohydrates concentration was
121	determined in triplicate by sulphuric-phenol method ^[22] using D-Glucose (Merck) as standard and
122	the same UV spectrophotometer against the blank at a wavelength of 485 nm. To convert into
123	COD, 1g carbohydrates assumed as $C_6H_{12}O_6$ is equivalent 1.07 g COD. ^[23] The coefficient of
124	variance was within 6.8% for ten identical samples. Soluble fractions of above-mentioned
125	parameters were obtained from by filtrating supernatant fraction of centrifuged sludge (10,000
126	rpm for 10 mins) through a 0.45 μ m membrane filter. Ammonia-Nitrogen was measured in
127	triplicate using Nessler method ^[18] by reading the absorbance at 425 nm. The COV was equal to
128	6.6% for ten identical samples. Soluble Phosphorus (as PO ₄ ³⁻) was analyzed using the
129	vanadomolybdophosphoric acid colorimetric method described in Standard Methods. ^[18] The
130	absorbance was read on the same spectrophotometer at 470 nm and the coefficient of variation for
131	three replicates was 0.6%. Particle size distribution was measured with particle size analyzer
132	(Shimadzu, model SALD-3101) according to laser diffraction. The median diameter was used to
133	quantify the particle size distribution. By definition it is the particle size such that 50% of the
134	particles are larger and 50% are smaller than that value.

Sludge disintegration degree (DD_{COD}) was used in this study to express the ratio of solubilized
COD to the maximum possible COD solubilization and can be used to quantify the sensitivities of
different sludge to ultrasound treatment: ^[24]

139
$$DD_{COD} = \frac{\text{SCOD}_{\text{T}} - \text{SCOD}_{\text{O}}}{\text{SCOD}_{\text{NaOH}} - \text{SCOD}_{\text{O}}}$$
 (1)

140 Where $SCOD_T$ is the Soluble COD of treated sample, $SCOD_{NaOH}$ is the Soluble COD of sample 141 immersed in 1M NaOH (ratio 1:1) at 90°C for 10 minutes and $SCOD_O$ is the soluble COD of the 142 raw sample.

143

144 Combined ultrasonic and thermal treatments

145

146 Experiments were carried out to investigate the effect of ultrasonication and thermal treatments 147 separately and in combination. The ultrasonication tests were carried out in batch mode using an 148 ultrasonicator (Misonix, Q700) with a frequency of 20 kHz and a maximum power input of 149 700W. A solid tip probe (#4208) with a diameter of 19.1 mm and maximum amplitude of 60 µm 150 was immersed 1-2 cm below the surface of the sludge. 80% of the maximum amplitude was used 151 and the corresponding power input was around 140W. Ultrasonication energy input was 152 quantified using both the ultrasonication time and the specific energy input calculated as follows: [16] 153

154
$$SEI = \frac{Total \, energy \, input \, (J)}{Total \, solids \, in \, sample \, (g)} = \frac{P * t}{V_{sludge} * TS}$$
 (2)

155 Where P is Power input of ultrasonicator (W), t is the time of ultrasonication (s), V_{sludge} is the 156 volume of treated sludge (L) and TS is the Total solids concentration of treated sludge (g/L). During ultrasonication the temperature was monitored and refrigerated below 30°C with ice-water
mixture if necessary.

159 For the combined treatment, ULS-treated sludge (at 5,000 kJ/kg TS unless stated otherwise) and

160 non treated sludge were mixed at a specific ratio in plastic capped tube (total volume was 10 mL)

161 and placed in a water bath (Haake DL30) without shaking for 24 hours to study the thermal

162 treatment. The temperatures tested for the combined ULS-thermal treatment were 25°C (ambient),

163 35°C, 45°C, 55°C, 65°C, 75°C and 85°C.

164

165 Anaerobic biodegradability

166

167 Prior to anaerobic biodegradability tests the sludge was pre-treated with ULS at 5,000 kJ/kg TS followed by thermal treatment at 65°C. Biochemical methane potential (BMP) assay were 168 conducted according to Owens et al.^[25] in 120 mL serum bottles to quantify the anaerobic 169 170 degradability of sludge samples. For the BMP assays 10 mL of substrate (raw or treated sludge), 171 20 mL of degassed acclimatized inoculum and 30 mL medium were added to serum bottles. 172 Mixture of 20% CO₂ and 80% N₂ were used to purge each bottle for three minutes to create 173 absolute anaerobic environment. Two blanks containing the inoculum and the medium were run in parallel, and the methane produced was subtracted from the methane produced in the bottles 174 175 containing the samples. All bottles were incubated in an orbital shaker at 35°C. The biogas 176 volumes were regularly measured using a wetted glass syringe and reported at atmospheric pressure and a temperature of 35°C.^[26] The composition of biogas was analyzed with gas 177chromatography (Agilent Technologies 7890A GC system) with two thermal conductivity 178179 detectors (TCD) and an Agilent HayeSep C 3.0 m X 1/8" X 2.0 mm packed column. The flow

180	rate was controlled at 45 mL/min and the temperatures of injector, oven and detector were 120°C,
181	115°C and 150°C, respectively.
182	
183	Results and discussion
184	
185	Preliminary tests on the effects of ultrasonication alone
186	
187	In this section the effects of ultrasonication alone were investigated. Various parameters such as
188	soluble carbohydrates, proteins, SCOD, phosphorus and ammonia are shown in Figure 1.
189	
190	[Figure 1]
191	
192	Figure 1A shows that ULS has a significant impact on soluble biopolymers with an increase in
193	SCOD, proteins and carbohydrates concentrations to 5.5 g/L, 1.6 g/L and 500 mg/L, respectively.
194	The increase in soluble carbohydrates was, relatively, less obvious because the sludge contained
195	thickened waste activated sludge which is rich in proteins from bacterial cells and EPS. Figure 1B
196	shows that ultrasonication had a significant effect on the soluble phosphorus concentration, which
197	means that ULS was able to break open the cells and release phospholipids from cell membrane
198	and phosphorus from the DNA into the bulk liquid. The concentration of ammonium in the
199	supernatant was also analyzed, and it was found that it slightly increased from 120 to 170 mg/L $$
200	during the first 5 minutes of treatment, but afterwards it remained constant. It is possible that

some proteins in the sludge were broken down or that ammonium from the cytoplasm wasreleased in the supernatant due to the action of ULS.

203

204 Figure 1D shows the evolution of various groups of particles based on their size. Cavitation 205 bubbles caused by ultrasound are known to disrupt floc structures and reduce floc size. Particles 206 larger than 100 µm or cells flocs and aggregates are readily disrupted by ULS within the first 207 minutes with the number of large flocs reduced from 26% to 12% and then below 5% as the SEI 208 reached 10,000 kJ/kg TS. At the same time, the number of colloidal particles or small flocs (13-209 100 µm) also dropped significantly due to physical disruption, while the amount of single cells, 210 small colonies and possibly cell debris (2-13 µm) started to increase markedly from 10% to 50%. This was consistent with Lehne et al.^[27] who found that an obvious floc size reduction took place 211 212 below a SEI of 3,000kJ/kg TS. Interestingly, beyond 10,000 kJ/kg TS higher SEIs became 213 inefficient. The slow and steady increase of intra-cellular materials and EPS (<2 µm) showed that 214 ULS could indeed disrupt cells walls and solubilise EPS even at low SEIs which was consistent 215 with the evolution of soluble biopolymers, ammonia and phosphorus. This contradicts Lehne et al. ^[27] who suggested that cell lysis did not take place until a SEI of 3,000 kJ/kg TS was applied. 216 217 Overall, it can be concluded that ULS was more efficient towards large flocs.

218

219 Preliminary tests on the combination of ULS and thermal treatments

220

In our preliminary tests it was observed that the temperature could increase up to 70°C during

222 ULS if a small volume of sample (<50 mL) was used and if the sample was not cooled down.

223 Using a pulse mode could reduce the heat generated, but was not efficient to solubilise more COD

224 (data not shown). It was then decided to investigate the effect of ULS and heat separately and by 225 combining ULS and thermal treatment in sequence. The sequence of thermal-ULS was tested and 226 it was found the thermal energy could lyze cells which then released soluble materials such as 227 colloids and proteins leading to an increase in SCOD concentration (data not shown). However, 228 during the subsequent ultrasonication the SCOD, soluble proteins and carbohydrates 229 concentration increased only by 700 mg/L, 60 mg/L and 145 mg/L, respectively, which was 230 deemed insignificant. It was postulated that the propagation of ultrasound waves was hindered 231 and could not reach intact cells. Therefore, we focused on the ULS-thermal sequence in this 232 study.

First, we ultrasonicated a specific percentage of the sludge (0-25-50-75-100%) and measured the SCOD concentration obtained after mixing with the non-treated fraction. The sludge was then incubated at a specific temperature without mixing and at neutral pH, and it can be seen in Figure 2 (top) that after 24 hours incubation at 30°C the SCOD increased to ~3 g/L for the 25% ULStreated sludge and decreased for the higher ratios presumably because SCOD was consumed by

238 mesophilic microorganisms at 30° C, and converted to CO₂.

239 Interestingly, the situation was very different at 55°C as shown in Figure 2 (bottom). When the

sludge was not ultrasonicated and placed at 55°C (100% raw) the SCOD increased to 5.35 g/L,

whereas with 25% ULS-treated sludge the SCOD increased to 7.1 g/L. The improvement of

solubilization of the sludge due to the combination of thermal and ultrasonication is in line with

243 previous studies, ^[28] but to our knowledge this is the first study where only a fraction of the

sludge is ultrasonicated during the combined ULS thermophilic pre-treatment.

245 Moreover, the use of ultrasonication on 25% of the sludge improved even further the performance

of thermal treatment. This was due to the disruption of flocs and the breakdown of cells

247 containing intra-cellular hydrolytic enzymes. This is in line with researchers who showed that enzymes could be extracted from WAS using ULS.^[29] However, it was demonstrated that the free 248 249 enzymatic activity present in the liquid phase was almost negligible, being immobilised on flocs 250 (connected to the polymeric extracellular substances) or attached to the cellular walls by ionic and hydrophobic interactions.^[30-31] Therefore, a physical treatment is useful to disrupt the flocs and 251 release the enzymes. Yu et al. ^[32] showed that ultrasonic pre-treatment enhanced enzymatic 252 253 activities and promoted the shifts of extracellular proteins, polysaccharides and enzymes from 254 inner layers of sludge flocs (i.e., pellet and tightly bound EPS) to outer layers (i.e., slime) and this 255 increased the contact and interaction among extracellular proteins, polysaccharides and enzymes 256 that were originally embedded in the sludge flocs, resulting in improved efficiency in the 257 subsequent aerobic degradation. 258 Heat and ultrasonication can also be used to rupture cells and release the intra-cellular proteases which can hydrolyze proteins in the sludge. Nabarlatz et al. ^[29] also found that the activity of 259 260 extracellular protease in activated sludge tank was much lower than that of intracellular protease, 261 therefore, it is sensible to use ULS prior to thermal treatment. At 100% ULS-treated sludge the 262 final SCOD concentration reached almost 8 g/L. However, at higher ratios of ULS-treated sludge

the improvement of ultrasonication became marginal.

264

265 [Figure 2]

266

267 Effect of temperature during the ULS-thermal tre-treatment of sludge

268

269

270 Based on the previous experiment a 25% ratio was used to determine the optimum temperature 271 for the enzymatic treatment. Figure 3 shows that the higher the temperature, the more COD was 272 solubilized (up to ~11 g/L). Higher SCOD (up to 14 g/L SCOD) could be obtained depending on 273 the initial solid concentration of the sludge (data not shown). Temperatures greater than 65°C 274 resulted in a marginal increase. It was also found that mixing during the thermophilic treatment 275 resulted in a 20% increase in SCOD concentration (data not shown). 276 To investigate further the effect of heat, a sample was autoclaved (121°C for 20 min) and the final 277 SCOD was only 6,700 mg/L. As the temperature rises slowly in an autoclave, the enzymes were 278 still active but could have been deactivated at high temperatures (>85°C) which limited the extent 279 of solubilization compared to a milder thermal process. This showed further that heat was not the 280 only phenomenon taking place. The solubilization of WAS by heat-treatment can be induced by sludge lysis and further cryptic growth (lysis-cryptic growth).^[33] In the lysis-cryptic growth, 281 282 sludge reduction is achieved because some portions of lysates are consumed for the catabolism and finally emitted as CO₂. This was confirmed using our sludge as a CO₂ production of 4.4 mL 283 284 and 6 mL was recorded after 1 hour incubation at 55°C and 65°C, respectively. After 24 hours, 285 the cumulative CO₂ production reached 9.9 mL and 10.2 mL, respectively, indicating the 286 consumption of SCOD for the growth of both thermophilic and hyper-thermophilic bacteria. 287

288 [Figure 3]

289

Yan et al. ^[34] used a simple heat-treatment process (700 ml was incubated at 60 °C, 120 rpm for
24 h in a 1 l Erlenmeyer flask) and also showed that there was rapid increase in population of
thermophilic bacteria at the early stage of heat-treatment and the emergence of protease-secreting

bacteria. Hasegawa et al. ^[12] showed that the hyper-thermophilic aerobic microbes were identified
as belonging to *Bacillus*.

Therefore, the potential for increased performance is inherent in the sludge itself,^[35] and although heat treatment is beneficial for solubilization, long thermal treatment are not interesting from a process point of view but also because some of the lyzate is consumed by thermophilic bacteria and lost as CO₂ and cannot be used to produce methane.

299

300 *Combination of different ratios of sludge treated by ULS prior to thermal pre-treatment at* 55°*C* 301

302 In this experiment a specific percentage of sludge (0, 5, 10, 20, 50 and 100%) was ultrasonicated, 303 then mixed it with the remaining non-ultrasonicated fraction and incubated in a water bath to study the kinetics of solubilization at 55°C and 65°C. As 75°C and 85°C were shown to result in a 304 305 marginal SCOD increase in the previous section, these temperatures were not tested further in 306 details in this study. Carbohydrates and proteins are two predominant biopolymers in EPS structure which also contributes a great part of COD to sludge.^[32] Therefore, the solubilization of 307 308 carbohydrates and proteins provide essential information about disintegration of sludge structure. 309 The soluble COD, proteins and carbohydrates concentrations obtained at 55°C are shown in 310 Figure 4.

311

It can be seen that the thermophilic treatment alone resulted in a final SCOD of 7.8 g/L, whereas a significant increase to 8 g/L, 8.7 and 9.3 g/L was observed when 20%, 50% and 100% of sludge was ultrasonicated prior to the thermal treatment, respectively. Below 20% of ULS-treated sludge there was a small effect as indicated by close SCOD values. The results indicated that as the

316	percentage of ULS increases, more cells are broken down and more intracellular materials are
317	released into the bulk as shown by higher SCOD concentrations. However, the effect of ULS was
318	not linear, meaning that 100% ULS treated did not result in twice the solubilization of 50% ULS-
319	treated sludge. This shows that treating 100% of sludge by ULS is not an interesting option,
320	however, 20% and above had a positive impact on the subsequent thermal treatment.
321	It was also found that ultrasonication increased the COD solubilization rate of the overall pre-
322	treatment. For instance, the thermophilic treatment took 24 hours to reach 7.8 g/L SCOD, whereas
323	only 3 hours thermal treatment was required when 100% sludge was ultrasonicated. This
324	demonstrated that the thermal treatment time can be significantly reduced by combining ULS.
325	
326	[Figure 4]
327	
328	It can be seen from Figure 4B and 4C ULS improved the rate of proteins and carbohydrates
329	solubilization compared to the thermophilic treatment alone. The concentration increased during
330	the first six hours of thermophilic treatment and decreased afterwards due to the consumption of
331	nitrogen and carbohydrates by thermophilic bacteria. Proteins and carbohydrates solubilization
332	might have continued after 6 hours, but it could not compensate for the uptake by opportunistic
333	thermophilic microorganisms, resulting in a net decrease after 6 hours of treatment. This net
334	decrease was, however, not observed in SCOD concentration (Fig. 4A) as COD analysis
335	encompassed various biopolymers including proteins and carbohydrates, and also lipids,
336	phosphates, ammonia, humic and fulvic acids that were solubilized.
337	

338 Combination of different ratios of sludge treated by ULS prior to thermal pre-treatment at 65°C 339

The soluble COD, proteins and carbohydrates concentrations obtained at 65°C are shown in 340 341 Figure 5. As expected, the extent and rates of COD, proteins and carbohydrates solubilization 342 was enhanced at 65°C compared to 55°C. This is due to improved cell lysis and possibly higher enzyme activity. Yu et al. ^[32] had indeed also showed that enzymatic activities (proteases, α -343 344 amylase, α -glucosidase, alkaline-phosphatase and acid-phosphatase) were markedly increased after ultrasonication. In terms of final SCOD concentration, 100% ULS was equivalent to 1 hour 345 346 hyper-thermophilic treatment. Both conditions resulted in ~5 g SCOD/L. When 100% of sludge 347 was ultrasonicated, less than 1 hr of hyper-thermophilic condition was required to reach 8 g 348 SCOD/L. However, 24 hrs were required to reach that level in individual hyper-thermophilic pre-349 treatment. Therefore, ULS shortened significantly the hyper-thermophilic treatment. Sahinkaya and Sevimli ^[28] reported that the SCOD increased from 55 to 3,500 mg/L after 10 minutes 350 351 ultrasonication (1.5 W/mL) at 80°C for 1 hour which was found to be the optimum temperature. 352 Ultrasonication alone resulted in a concentration of 2,250 mg/L. This confirmed the better results 353 using a combination of ultrasound and thermal treatments. However, these concentrations were 354 much lower than in this study due to lower TS level (4 g/L) in the raw sludge.

355

356 [Figure 5]

357

358 It can be seen that the extent of protein solubilization increased as the percentage of ULS-treated 359 sludge increased. This is in line with the previous observations at 55°C. However, at 65°C, the 360 effect of ULS was more dominant as shown by a significantly higher solubilization rate at

361 percentages as low as 10%. Interestingly, there was no net decrease in soluble proteins 362 concentrations at 65°C in contrast to what was observed at 55°C. This indicates that the rate of 363 proteins solubilization was higher than the rate of proteins degradation and consumption by 364 hyper-thermophilic bacteria. Soluble carbohydrates, however, were consumed by hyper-365 thermophilic bacteria as indicated by a net decrease in concentration after 6 hours. The existence 366 of such hyper-thermophilic bacteria was previously documented and was found to belong to *Bacillus*.^[12]The net decrease was insignificant for the 100% ULS-treated sample showing that 367 368 ULS could also inhibit to some extent the growth of the hyper-thermophilic bacteria and avoid the 369 consumption of soluble carbohydrates.

370

371 TSS and VSS removals during ULS and thermal pre-treatment

372

373 [Table shows the TSS and VSS removal during the individual and combined pre-treatments. It 374 can be seen that ULS alone resulted in TSS removals lower than 10%, while the thermal treatment 375 resulted in TSS removals in the range 20-23%. When 50% of the sludge was ultrasonicated and 376 treated at 65°C, then a maximum of 27% TSS and VSS removal was obtained. Treating 100% of 377 the sludge by ultrasonication did not increase this removal, confirming that ultrasonication of a 378 high proportion is not required. Yu et al. ^[32] obtained 11.8% TSS reduction after an ultrasonication treatment (10 min, 3 kW/L) of 379 380 WAS and attributed this to the release of soluble organic carbon sources and extracellular 381 enzymes, and the enhanced contact between them. The sludge reduction for TSS was 30.9% after aerobic degradation (compared with 20.9% in the control) showing that the ultrasonic pre-382

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388	
389	[Table 3]
390	Effect of the combined pre-treatment on anaerobic biodegradability
391	
392	The effect of ULS alone on the anaerobic biodegradability was tested first. As shown in Figure 6
393	(a), the biodegradability of all the ultrasonicated sludge was higher than the control. The ultimate
394	biodegradability increased with increasing specific energy input. However, ultrasonication at high
395	specific energy input may not be economical. It was found that 9 kJ/g TS ultrasonication
396	improved the sludge biodegradability by 14.8%, whereas at a SEI 7 times higher (i.e. 63 kJ/g TS),
397	the biodegradability increased by 31.8% which was only 2.2 times higher compared to the
398	improvement induced by 9 kJ/g TS ultrasonication.
399	In order to evaluate the possibility to use SCOD data to predict the biodegradability, the sludge
400	ultimate biodegradability and SCOD concentrations were plotted in Figure 6 (b). Linear
401	regression was found to be the most suitable model to describe the relationship. The coefficient of
402	determination (R^2) was 94.83%, indicating a strong correlation which is in line with Bougrier et
403	al. [36] who observed that biogas increase in ultrasonicated sludge originated mainly from the
404	soluble fraction.
405	

treatment could significantly enhance aerobic digestion efficiency and the extent of sludge

383

384

biodegradability.

407	Since it was found that higher SCOD were obtained by combining ULS and thermal treatment (up
408	to 14 g/L), a higher methane production was expected to be found using the combined pre-
409	treatment. Several combinations of pre-treatments were tested and the BMP results are shown in
410	Figure 7. A small percentage (5%,~5,000 kJ/kg TS) of ULS-treated WAS was combined with the
411	thermal treatment at 65°C for 24 hours and it was found that the methane production increased by
412	20%. This was higher than previous studies ^[28] where 13.6% increase in methane was obtained
413	after 1 minute ultrasonication (1 W/mL) and 1 hour thermal treatment at 80°C. It was also found
414	that methane production was greater with the combination compared to ultrasonication of 5% or
415	even 100% alone. The methane percentage in the biogas was up to 6% higher indicating a higher
416	calorific value due to the combined pre-treatment. However, a lag-phase of 8-12 days was
417	observed following the combined treatment which may be the result of a higher SCOD and its
418	components which the anaerobic inoculum was not acclimated to. Gavala et al. [37] found that
419	there are indigenous microorganisms in primary sludge capable of methane production and
420	incubation at 70°C for 1 day or more as a pre-treatment resulted in their inactivation.
421	Furthermore, they found that the thermal pre-treatment of both primary and secondary sludges led
422	to increased hydrogen levels that can inhibit methane production. ^[26] Our anaerobic inoculum
423	may have not contained enough hydrogenotrophic species which led to some inhibition and the
424	observed lag-phase.
425	Moreover, this combination (+20%) was more efficient that ULS alone at high SEI (100%, 9,000
426	kJ/kg TS) as shown in Figure 6(a) (+14.8%). This is due to the COD solubilization obtained after
427	the pre-treatment. After 100% ULS typical SCOD concentrations are in the range 4-5 g/L (Fig.
428	1A), whereas the combination of ULS and thermal treatment resulted in 10-11 g/L SCOD.

429	ULS is a fast method, but relatively inefficient to solubilise COD and it is expensive to treat
430	100% of WAS. Thermal treatment is efficient to solubilise COD, but is a slow process. Thermal
431	treatment could be a viable option to consider if waste heat is available on site. It was found that
432	these disadvantages can be alleviated when both methods are combined together, while methane
433	production is improved. Further work is needed to find an optimum combination.
434	

435

437 Conclusion

[Figure 7]

438

439 In this paper we investigated the pre-treatment of sewage sludge using ultrasonic and thermal 440 treatments. Ultrasonication had a marked effect on particles with size greater than 100 microns 441 (flocs) and in the range 13-100 microns (cells, colonies or colloids) at specific energy input lower 442 than 10,000 kJ/kg TS. The optimum temperature during the thermal treatment was found to be 443 65°C. It was found the combination of ULS (30 sec., 5,000 kJ/kg TS) and thermal treatments 444 resulted in greater solubilization of COD (760 to 10,200 mg/L), proteins (115 to 2,900 mg/L) and 445 carbohydrates (60 to 660 mg/L) than individual treatments. During ultrasonication treatment 446 alone (30 sec., 5,000 kJ/kg TS), SCOD, soluble proteins and carbohydrates concentrations 447 increased to 4,700 mg/L, 1,000 mg/L and 500 mg/L, respectively. The ultrasonication of 50% of 448 the sludge followed by the incubation at 65°C could increase the SCOD from 760 mg/L to 9,300 449 mg/L. It was also found that ultrasonication increased the COD solubilization rate of the subsequent thermal treatment at 65°C and treatment time could then be reduced to a few hours (1-450 6 hours) instead of 24 hours or several days. The SCOD obtained after ultrasonication pre-451

452	treatment and its anaerobic biodegradability was found to be linearly correlated ($R^2 = 95\%$). The
453	combined treatment resulted in 20 % increase in biogas production.
454	
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456	
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584	FIGURE CAPTIONS
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586	Figure 1. Effect of ultrasonication: Specific Energy Input on (A) soluble proteins, carbohydrates
587	and COD, (B) soluble phosphorus concentration, (C) soluble ammonia concentration, (D)
588	evolution of various groups of particles based on the size. The ammonia and phosphorus
589	concentration remained constant beyond 6,000 kJ/kg TS.
590	
591	Figure 2. Effect of ultrasonication of 0, 25, 50, 75 and 100% of sludge (30 sec, ~5000 kJ/kg TS)
592	followed by thermal treatment at 30° C (top) and 55° C (bottom).
593	
594	Figure 3. Effect of the incubation temperature on the SCOD during the thermal treatment
595	following ultrasonication of 25% of sludge (~ 5,000 kJ/kg TS).
596	
597	Figure 4. Evolution with time of the SCOD (A), soluble proteins (B) and carbohydrates (C)
598	during the thermal treatment at 55°C following the ultrasonication of 0, 5, 10, 20, 50 and 100% of
599	sludge (~ 5,000 kJ/kg TS).
600	

601	Figure 5. Evolution with time of the SCOD (A), soluble proteins (B) and carbohydrates (C)
602	during the thermal treatment at 65°C following the ultrasonication of 0, 5, 10, 20, 50 and 100% of
603	sludge (~ 5,000 kJ/kg TS).
604	
605	Figure 6. (a) Anaerobic digestion tests of control and ultrasonicated sludge (b) Linear fitting of
606	sludge biodegradability and SCOD concentration after ULS pre-treatment.
607	
608	Figure 7. Cumulative methane production after several combinations of ULS and thermal pre-
609	treatments.
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612	TABLE CAPTIONS
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615	Table 1. thermal pre-treatment methods
616	
617	Table 2. Properties of the sewage sludge used in this study. NM= not measured.
618	
619	Table 3. TSS and VSS removal during the combined ULS/thermal pre-treatment.
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- 630 Fig. 1

- 639

















- 690 Fig. 5





	Treatment conditions	Anaerobic digestion Conditions	Results	Reference
	Microaerobic, 60–70°C, 1	Batch, 10 days 37°C	Increase of biogas production from 200 to 300 mL.g ⁻¹ VS _{fed} (+50%)	[37]
	day Microaerobic 65°C, 1 day	CSTR, HRT: 21 and 42 days 35°C	Increase of COD removal (+30%) No methane production increase	[38]
	70°C	Batch	Increase of CH ₄ production from 8.30	[37]
	7 days 70°C	37°C Batch	to 10.45 mmol.g ⁻¹ VS _{fed} (+26%) CH ₄ production of 10.9 mmol.g ⁻¹	[37]
	7 days 70°C	55°C Batch	VS _{fed} (no influence) Increase of CH ₄ production from 21.2	[37]
	4 days 70°C	37°C Batch	to 24.7mmol g^{-1} VS _{fed} (+16%) Increase of CH ₄ production from 13.7	[37]
	7 days 70°C 2 days	55°C CSTR, HRT: 13 days (15 days without	to 25.5 mmol.g ⁻¹ VS _{fed} (+86%) Increase of CH ₄ production from 40 to 55 mL.L ⁻¹ d ⁻¹ (+28%)	[39]
	70°C 9, 24, 48 h	pretreatment) 55°C CSTR, HRT: 10 days 55°C	Increase of CH ₄ production from 0.15 to 0.18 mL.g ⁻¹ VS _{fed} (+20%) Increase of energy production (+60–100%)	[40-41]
	70°C 2 days	CSTR, HRT: 13 days (15 days without pretreatment)	Increase of CH ₄ production from 13.6 to 20.1 mmol.g ⁻¹ VS _{fed} (+48%)	[17]
	50–65°C 2 days	CSTR HRT: 13–14 days 35°C	Increase of CH ₄ production (+25%) compared to 35°C pretreatment	[42]
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Table 1. thermal pre-treatment methods

Table 2. Properties of the sewage sludge used in this study.

	Parameters (acronym, unit)	WAS	Anaerobic Inoculum
	рН	5.9-6	7.3
	Soluble Chemical Oxygen Demand (SCOD, mg/L)	670 - 1440	454 ± 8
	Total Chemical Oxygen Demand	18 - 25	13.75 ± 0.53
	(1COD, g/L)	126 170	0.5 ± 0.2
	$\frac{101a1}{2} \text{ Solids} (15, g/L)$	13.0 -17.2	9.5 ± 0.5 7.1 + 0.2
	Total Supported Solida (TSS of L)	10.7 - 15.4 12.4 15.0	7.1 ± 0.3
	Volatila Suspended Solids (VSS, g/L)	12.4 - 13.9	9.5 ± 0.2
	Ammonia (mg N/L)	10.3 - 13.0 122.07 ± 2.72	7 ± 0.3
	Phosphate (mg PO_4^{3-}/I)	122.97 ± 2.72 24.11 ± 4.71	NM
733	NM- not measured	24.11 - 4.71	1 1 1 1 1 1
734	NM – not measured.		
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755	Table 3	TSS and VSS	romoval	Juring the	combined	III S/thormol	nra traatmant
155	Table 5.	. 155 and v55	removal	Juring the	combined	ULS/merman	pre-treatment.

		TSS removal %	VSS removal %
	Raw	0	0
	ULS 20% (5,000 kJ/kg TS)	4.6	2.36
	ULS 50% (5,000 kJ/kg TS)	7.28	5.91
	ULS 100% (5,000 kJ/kg TS)	8.62	6.86
	raw+55°C for 24hrs	20.5	19.15
	raw+65°C for 24hrs	22.22	22.93
	ULS 20% + 55°C for 24hrs	21.65	23.4
	ULS 50% + 55°C for 24hrs	21.46	20.57
	ULS 100% + 55°C for 24hrs	22.8	22.46
	ULS 20% + 65°C for 24hrs	23.75	23.4
	ULS 50% + 65°C for 24hrs	27.2	26.95
756 757 758 759 760 761 762 763 764 765 766	ULS 100% + 65°C for 24hrs	24.33	24.35
767 768			

SUPPORTING INFORMATION

Figure 1S. (A) Samples (1 µL) taken after 6 hours of hyper-thermophilic pre-treatment at 65°C with and without ultrasonication and pipetted into wells on Petri dishes placed at 55°C. The Petri dishes contain several replicated wells. (B) Samples taken after 6 hours of enzymatic pre-treatment at 65°C with and without ultrasonication and pipetted into wells on Petri dishes placed at 37°C. (C) Samples taken after 24 hours of hyper-thermophilic enzymatic pre-treatment at 65°C with and without ultrasonication and pipetted into wells on Petri dishes placed at 37°C.





WAS: 100 % ULS, 6 h at 65°C

Petri dish: 24 hrs at 55°C

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A WAS: 0 % ULS, 6 h at 65°C Petri dish: 24 hrs at 55°C



B WAS: 0 % ULS, 6 h at 65°C Petri dish: 96 hrs at 37°C



WAS: 100 % ULS, 6 h at 65°C Petri dish: 96 hrs at 37°C



WAS: 100 % ULS, 24 h at 65°C Petri dish: 96 hrs at 37°C



