

RESEARCH ARTICLE

Accumulating the key proteomic signatures associated with delirium: Evidence from systematic review

Md Parvez Mosharaf^{1,2*}, Khorshed Alam¹, Jeff Gow^{1,3}, Rashidul Alam Mahumud⁴

1 School of Business, Faculty of Business, Education, Law and Arts, University of Southern Queensland, Toowoomba, Queensland, Australia, **2** Bioinformatics Lab, Department of Statistics, University of Rajshahi, Rajshahi, Bangladesh, **3** School of Accounting, Economics and Finance, University of KwaZulu-Natal, Durban, South Africa, **4** NHMRC Clinical Trials Centre, Faculty of Medicine and Health, The University of Sydney, Camperdown, New South Wales, Australia

* parvez.mosharaf@unisq.edu.au



OPEN ACCESS

Citation: Mosharaf MP, Alam K, Gow J, Mahumud RA (2024) Accumulating the key proteomic signatures associated with delirium: Evidence from systematic review. PLoS ONE 19(12): e0309827. <https://doi.org/10.1371/journal.pone.0309827>

Editor: Hikaru Hori, Fukuoka University, JAPAN

Received: February 29, 2024

Accepted: August 12, 2024

Published: December 19, 2024

Copyright: © 2024 Mosharaf et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the article and its [Supporting information files](#).

Funding: This work was completed as a component of the first author's PhD study at the University of Southern Queensland in Australia, which was made possible by funding from the Australian Government Research Training Program Scholarship. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript".

Competing interests: The authors have declared that no competing interests exist.

Abstract

Delirium is a severe neuropsychiatric illness that occurs frequently in intensive care and postoperative units which results in prolonged hospital stays and increases patient's mortality and morbidity rates. This review focused on accumulating the common key proteomic signatures significantly associated with delirium. We carried out a systematic literature review of studies on delirium proteomic biomarkers published between 1st January 2000 and 31st December 2023 from the following electronic bibliographic databases including PubMed, Scopus, and EBSCOhost (CINAHL, Medline). A total of 1746 studies were identified and reviewed, and 78 studies were included in our review. The PRISMA guidelines, the PEO framework, and JBI quality assessment method were followed in this review to maintain the inclusion and exclusion criteria and risk of bias assessment. Most of the included studies were of the cohort (68%) and case-control (23%) design. We have accumulated a total of 313 proteins or gene encoded proteins of which 189 were unique. Among the unique proteins, we focused on the top 13 most investigated proteins (IL-6, CRP, IL-8, S100B, IL-10, TNF- α , IL-1 β , Cortisol, MCP-1, GFAP, IGF-1, IL-1 α , and NFL) that are significantly associated with delirium. Most of these are cytokines and inflammatory proteins indicating a strong interconnection with delirium. There was remarkable inconsistency among the studies in reporting the specific potential proteomic biomarker. No single proteomic biomarker can be solely used to diagnose and predict delirium. The current review provides a rationale for further molecular investigation of delirium-related proteomic biomarkers. Also, it's recommended to conduct further in-depth molecular research to decipher drug target biomolecules for potential prognostic, diagnostic, and therapeutic development against delirium.

Introduction

Delirium is regarded as a multifactorial medical condition, and its underlying pathologies might be caused by trauma, stress, or inflammation. Delirium is a severe but treatable medical

disorder that has been known for more than 2500 years. More than 30 terms have been used to describe it, including disturbance in attention and consciousness which tends to oscillate for a short term [1, 2]. Delirium is often poorly diagnosed and remained largely unrecognized among hospitalized patients, particularly in intensive care units (ICU) [3–5]. Delirium in the elderly is becoming more common, affecting up to 50% of adult hospitalized patients [6–8]. Delirium has a significant impact on a patient's recovery and increases complications in hospital settings, which extend hospital stays, raise overall costs, and increases mortality [9]. The three main hypotheses for delirium development and its progression include the alteration of neurotransmitter systems, the activity of inflammatory cytokines leading to permeabilization of the blood-brain barrier, and disruption of the hypothalamic-pituitary axis in response to severe trauma [10].

Yet, delirious patients in ICU/hospital settings may benefit from additional biological, molecular, and pathophysiological insights provided by molecular biomarkers associated with delirium incidence [11]. Genetic biomarkers are mainly classified into three basic groups: risk markers, disease markers, and end-products. The related biomarkers of delirium have been identified by several systematic reviews, and these biomarkers include distinct cerebrospinal fluid, amino acids, proteins, genes, regulatory molecules, genetic variation (i.e., SNIP), and other molecules as well [12–14]. Despite some discrepancies in the results, the identified biomarkers thus so far are internally linked by known functional interactions and molecular pathways [12]. Over the past few decades, the complexity of the molecular network-based biological functions and pathomechanisms influencing delirium development and its severity have been identified. There remains a knowledge gap about genetic factors, their regulatory elements, functional and molecular pathways, and the pathomechanisms of delirium origin and progression.

It has been observed that the pathophysiology of delirium and its complications in medical settings remain unknown based on the body of existing literature [15]. The molecular investigation is one of the effective and modern techniques that may assist with diagnosis, evaluation, and treatment while also shedding light on its mysterious pathogenesis [11, 16, 17]. In this aspect, the proteomic biomarkers efficiently indicate the severity, risk, onset, and recovery of the disease and disease motion. They can be treated as a potential therapeutic target for drug development [18, 19]. Even though delirium has been linked to certain biomarkers, research has revealed conflicting results, leaving no clear biomarkers for delirium. Yet, only a few studies have been conducted to accumulate the molecular proteomic biomarkers of delirium, indicating a lack of knowledge about this critical medical condition.

Therefore, this study focused on accumulating and identifying the key common proteomic signatures associated with delirium that have been studied so far. The review also justified the proteomic functional diversity of the common proteins associated with delirium. In addition, we have provided a comprehensive summary of the current state of knowledge on the proteomic signatures of delirium, which may form the basis of future in-depth molecular research and ultimately help with the development of more effective and potent drugs for delirium treatment.

Materials and methods

Systematic review

We conducted a systematic analysis of the literature to identify research on delirium-associated proteomic biomarkers. The entire procedure was guided by the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) standards, [20] and associated

PRISMA flowchart. The search strategy, inclusion and exclusion criteria utilized the PEO framework described below [21]:

Population: The study population solely included confirmed cases of delirium in humans. **Exposure:** The delirium-associated proteins/gene-encoded proteins which are the biomarker proteins that are significantly associated with delirium. **Outcome:**—this was delirium, and the identification of significant proteomic biomarkers associated with delirium. **Study design—**In the systematic review, studies from all kinds of observational and experimental were considered. This review was registered on PROSPERO (registration number: CRD42024566515).

Search strategy

A comprehensive electronic literature search was conducted on the selected electronic bibliographic databases (PubMed, Scopus, and EBSCOhost (CINAHL, MEDLINE)) using the MeSH terms, keywords, and subject headings. Only studies that were published in journals between 1st January 2000 and 31st December 2023 were considered for screening. The primary keywords were “delirium” and “biomarkers” used along with a combination of other associated keywords including “markers”, ‘genetics”, “genes” and “proteins” to search the studies. Boolean operators “AND”, and “OR” were applied to combine the searching keywords. In addition, this review was complemented by a thorough manual search of related studies. Further, studies were identified through citation searches of included studies and manual searches for professional web sources and key journals in these fields of research. The details of the search sentences used in different databases and their search outcomes are provided in [S1 File](#).

Eligibility criteria

Eligible studies were included if they were i) based on original research studies focused and reported on genes/proteins showing any statistically significant relationship between delirium and genes/proteins in human cases; ii) delirium was assessed and confirmed using established delirium assessment methods; and iii) were published between January 1, 2000, and December 31, 2023, in English. Otherwise, editorials, letters, perspectives, commentaries, reports, reviews and meta-analysis, study protocols, publications in other languages, and studies with ‘insufficient related data’ were excluded.

Study screening and selection process

The eligibility of studies to be included was determined following a three-stage screening process. The first stage involved screening of studies by title to eliminate duplications. The second stage required reading abstracts to determine their relevance to our study. Finally, the third stage necessitated reading full texts of the retained studies, and those that met the set criteria were kept. After screening the title and abstract, MPM and RAM carried out the full text screening to select the articles. During this process, we have discussed and reached a consensus with the other authors (KA and JG) to resolve any discrepancies.

Quality assessment

Quality assessment of the 78 included studies was conducted because of the heterogeneity among the study designs of the included studies. In this systematic review, cohort, case-control, cross-sectional, randomized control trials, and longitudinal study designs were found among the included studies. The Joanna Briggs Institute (JBI) [22] provided critical quality

assessment tools that have been utilized in this study for quality assessment. The JBI quality appraisal tools are widely used in academic studies to assess the risk of bias (graded as high, moderate, or low) [23–26] where the higher quality scores demonstrate better confidence and vice versa. The JBI appraisal tool was used to evaluate the 53 cohort studies, 18 case-controls, three randomized control trials, two cross-sectional, and two longitudinal studies included in our review. The overall quality appraisal scores are summarized in [S2 File](#).

Data extraction

The data were extracted from Mendeley libraries by one researcher (MPM) with the direct help and guidance of RAM, who subsequently reviewed the results. Discrepancies in the data were addressed and resolved by consensus, and in cases where the two researchers could not reach a consensus, other researchers (KA and JG) were consulted for adjudication. The studies reported the genes and proteins significantly related to delirium were mainly considered for this qualitative synthesis. At the data extraction stage from the selected articles, we considered the first author, publication year, age, gender, data collection time, method of detecting proteins/genes, country of study, study design, method of delirium assessment, and the reported significant proteins associated with delirium. Any missing information was kept blank or noted by “NA” on the data extraction table. The entire procedure was guided and completed by the systematic literature review tool Covidence (<https://app.covidence.org>).

Results

Description of included studies

Our search identified 1746 records, of which 1365 were screened ([Fig 1](#)). After removing 381 duplicates, 1365 records were excluded at title and abstract screening stage including for example, review papers, editorials, protocols, irrelevant and insufficient information, and others. Finally, 133 full-text studies were assessed for eligibility, and 78 studies were included in the systematic literature review ([Fig 1](#)). [Fig 1](#) demonstrates the PRISMA flowchart and the PRISMA Checklist has been provided in [S3 File](#).

The included studies differed significantly in terms of patients, research methodology and settings, and the biomarker investigated ([Fig 2](#)). In terms of context, participants were from either a medical or surgical environment setting. In the delirium-only trials, authors either did not include patients with comorbidities or did not assess neurocognition to ascertain whether comorbidities were present or not. Studies with additional comorbidities did not consistently account for these variables' existence. Pre-existing cognitive impairment and Alzheimer's disease were considered as dementia in this study and hence excluded.

All 78 studies have utilized different well-established delirium assessment methods to identify delirium either in the pre-operative or postoperative stage for critically ill patients. Twelve different delirium assessment methods were used by the included studies. Among them most of the studies used the Confusion Assessment Method (CAM; $n = 35$ studies) or its imitation for ICU (CAM-ICU; $n = 25$ studies) for delirium screening ([Fig 2A](#)). Other studies utilized the Diagnostic and Statistical Manual of Mental Disorders, 4th edition (DSM-IV; $n = 4$ studies) and Delirium Observation Scale (DOS), Delirium Rating Scale-Revised-98 (DRSR-98) and Nursing Delirium Screening Scale (NuDesc) were utilized by three single studies individually. A few studies were also utilized multiple assessment method to diagnose delirium ([Fig 2A](#)). Among the selected studies, most of them were conducted in the USA ($n = 22$ studies), followed by China ($n = 17$ studies), the Netherlands ($n = 10$ studies), the Germany ($n = 6$ studies), the Poland ($n = 5$ studies), the India ($n = 2$ studies) and the Norway ($n = 2$ studies), and other 11 different countries ([Fig 2B](#)). The included studies have a diversity of study settings

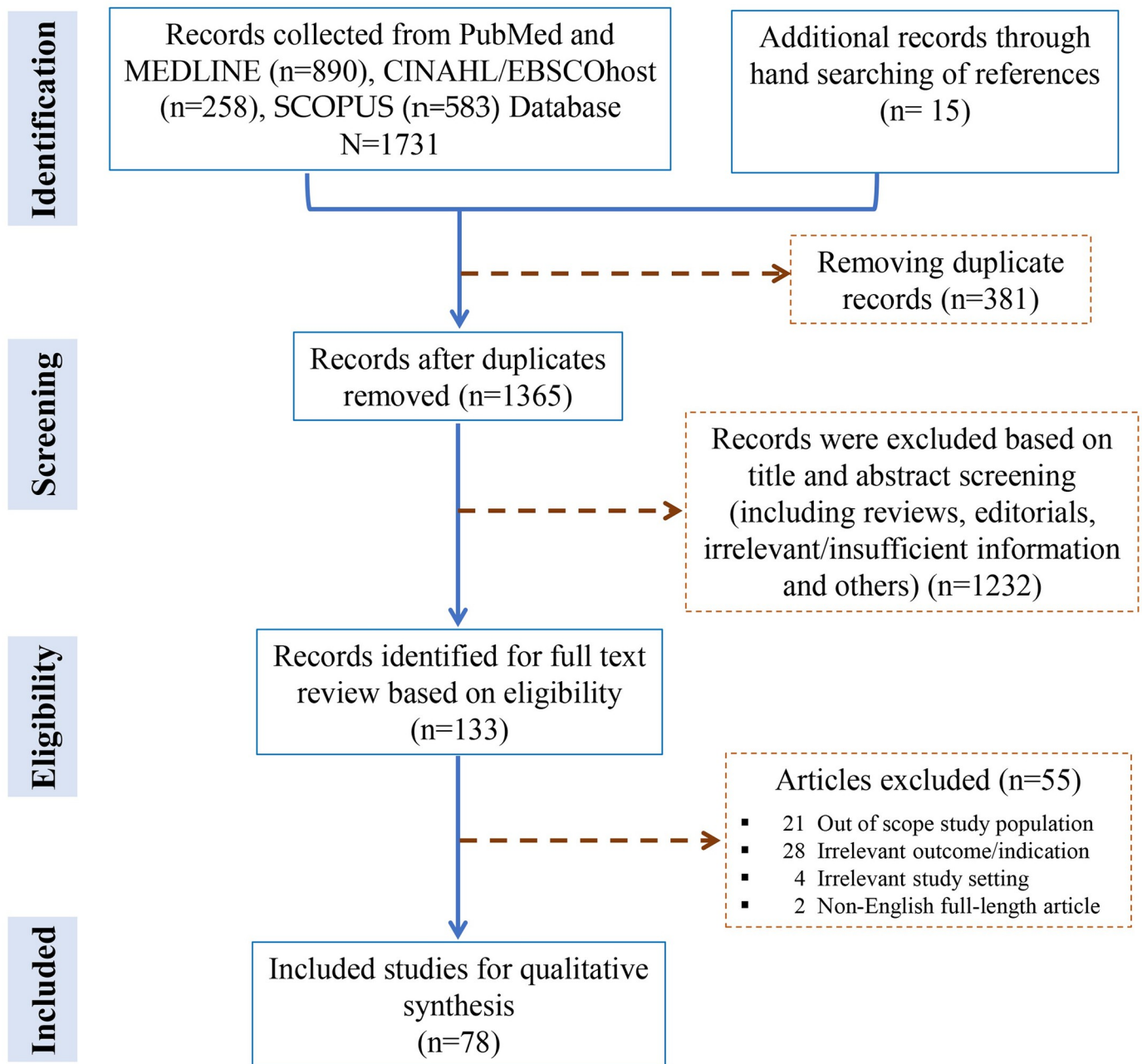


Fig 1. The PRISMA flow diagram of this study.

<https://doi.org/10.1371/journal.pone.0309827.g001>

including n = 53 (68%) cohort studies, n = 18 (23%) case-control studies, and other study settings as well (Table 1 and Fig 2C). The patient's demographic and study characteristics, including age, gender, sample size, number of delirious cases, and methods used to identify proteins/genes, have been documented in S1 Table. The screened full-text studies with decision, entire data matrix have been recorded in S2 Table. The authors' name and publication year, country of the study, type of study, delirium assessment method and the reported proteins/gene encoded proteins have been summarized in Table 1.

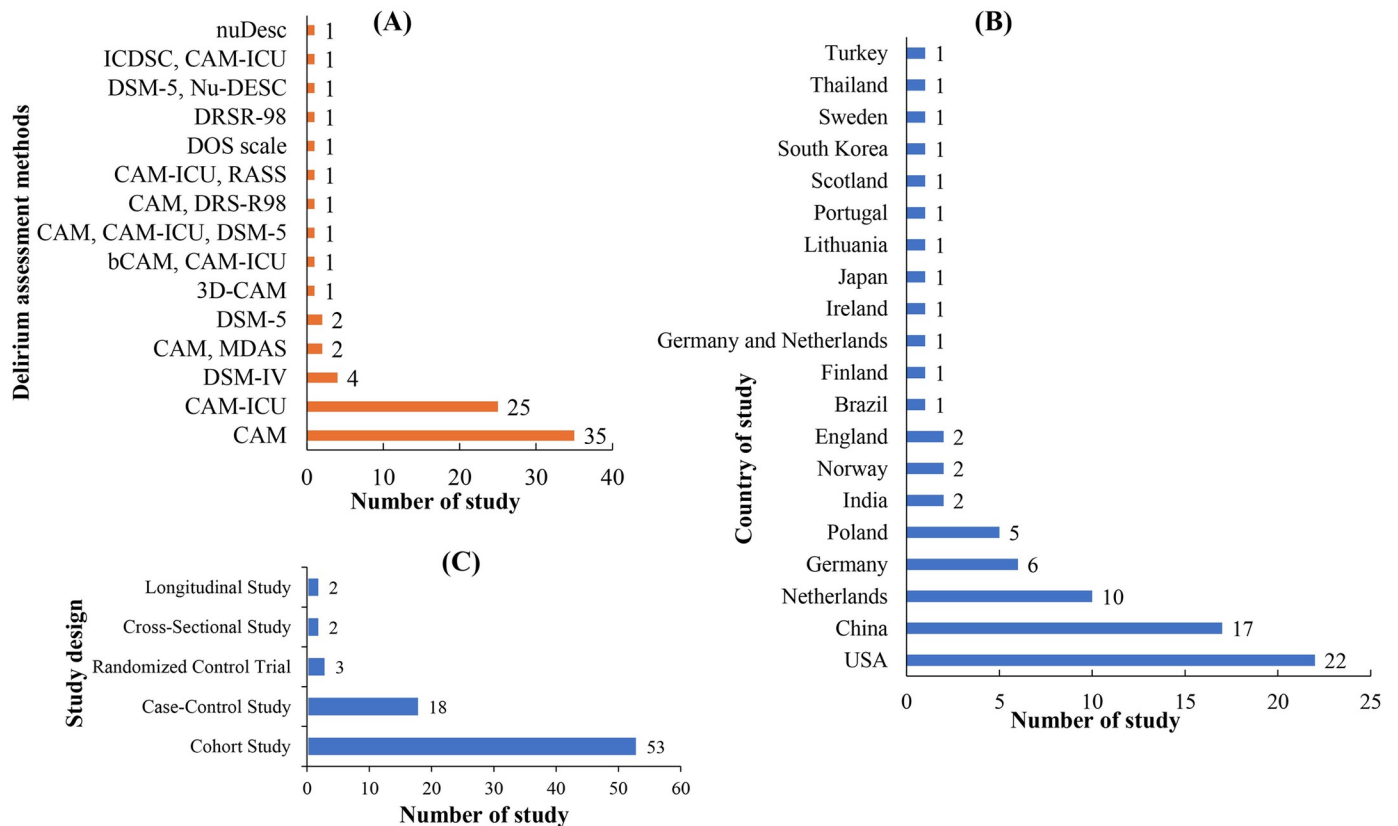


Fig 2. Distribution of the included studies according to (A) the delirium assessment method, (B) study design, and (C) country of study. [Here, (b)(3D)CAM: (Brief) (3 minutes) Confusion Assessment Method; CAM-ICU: Confusion Assessment Method for ICU; DSM-IV and DSM-5: Diagnostic and Statistical Manual of Mental Disorders, 4th and 5th edition; DOS Scale: Delirium Observation Scale, DRSR-98/DRS-R98: Delirium Rating Scale-Revised-98; NuDesc: Nursing Delirium Screening Scale; ICDSC: Intensive Care Delirium Screening Checklist; RASS: Richmond Agitation Sedation Scale; MDAS: Memorial Delirium Assessment Scale].

<https://doi.org/10.1371/journal.pone.0309827.g002>

Quality of the included studies

The JBI quality appraisal checklists provided the quality scores for the included studies (S2 File). The majority of the included studies were of medium/moderate quality ($n = 55/78$, 70.5%) and high quality ($n = 23/78$, 29.5%), indicating the robustness of the included studies (S2 File). For instance, among the cohort studies ($n = 53$), 47 studies were assessed as medium quality on the JBI scale when six were of high quality. Based on the quality appraisal checklists, 11 case-control studies were of high quality and seven were of the medium; among the RCT, two studies were of high, and one study was of medium quality. The two cross-sectional and two longitudinal studies were of high quality. In this review, no study was disqualified because of receiving a low-quality rating.

Delirium-associated important biomolecules

The included studies showed a total of 313 delirium-associated gene-encoded proteins where there were 189 unique proteins (Table 1 and S4 File). A few proteins were examined repeatedly in a substantial number of studies (Fig 3).

The most studied 13 proteins out of the reported proteins collected from the included studies have been highlighted (Fig 3) in this review as the key common proteins associated with

Table 1. Summary information of the 78 selected studies.

Authors & year	Country	Type of Study	Delirium Assessment Methods	All Associated Proteins/Genes
Adamis et al, 2009 [27]	Ireland	Longitudinal Study	CAM	IGF-1, IL-1ra, IFN- γ
Egberts et al, 2015 [28]	Netherlands	Case-Control Study	DSM-IV	Neopterin, IL-6, IGF-1
Hirsch et al, 2016 [29]	USA	Cohort Study	CAM	IL-5, IL-6, RAGE, IL-8, MCP-1, IL-10, IFN-a, IL-4, IFN- γ , IL-12, TNF-a, MIP-1 α , MIP-1 β
Kazmierski J et al, 2014 [30]	Poland	Cohort Study	CAM-ICU	IL-2, TNF-a
Miao et al, 2018 [31]	China	Cohort Study	DSM-IV	Neopterin, CRP, IL-6, IGF-1
Ritter et al, 2014 [32]	Brazil	Cohort Study	CAM-ICU	STNFR1, STNFR2, adiponectin, IL-1b
Sun et al, 2016 [33]	China	Cohort Study	CAM	IL-6, CRP, procalcitonin, Cortisol, ABI-40
Van Munster et al, 2008 [34]	Netherlands	Cohort Study	CAM	IL-6, IL-8
Vasunilashorn SM et al, 2015 [35]	USA	Case-Control Study	CAM	IL-6, IL-2, TNF-a, IL-12, VEGF
Liu X et al, 2023 [36]	China	Case-Control Study	CAM-ICU	NFL, GFAP
Sun Y et al, 2023 [37]	China	Cohort Study	CAM	CRP
Oren RL et al, 2023 [38]	USA	Cohort Study	3D-CAM	IL-8, LTBR, IL-6, ASGR1
Wu X et al, 2023a [39]	China	Cohort Study	CAM	A β 40, A β 42, P-Tau
Ruhnau J et al, 2023 [40]	Germany	Cohort Study	DSM-5, Nu-DESC	sTREM2, Gasdermin D, IL-6, S100B, IL-1B
Westhoff et al, 2013 [41]	Netherlands	Cohort Study	CAM	IL-6, IL-1ra, FLT-31
Heinrich M et al, 2021 [42]	Germany and Netherlands	Cohort Study	CAM	CHRM2, CHRM4
Van Munster et al, 2010 [43]	Netherlands	Cohort Study	CAM	DRD2, DRD3, SLC6A3
Terrelonge M et al, 2022 [44]	USA	Case-Control Study	CAM	FKBP5, KIBRA, KLOTTHO, MTNR1B, SIRT1
Yamanashi T et al, 2021 [45]	USA	Cohort Study	CAM-ICU	TNF
Yamanashi T et al, 2021 [46]	USA	Cohort Study	CAM-ICU	TNF-a, IL-1b, IL-6
Steimer M et al, 2021 [47]	Germany	Cohort Study	nuDesc	PER2, HO1
Nekrosius D et al, 2019 [48]	Lithuania	Cohort Study	CAM	COMT
Rhee J et al, 2021 [49]	USA	Randomized Control Trial	CAM	FN1.4, FN1.3, Troponin-1, C5a, IL-1, Cadherin-12, IL-6, PKC-Z, FGF-16, TIMP-1
Ballweg T et al, 2021 [50]	USA	Cohort Study	CAM-ICU	IL-8, IL-10, MCP-1
Tang C et al, 2020 [51]	China	Randomized Control Trial	CAM	IL-6, TNF-a, IL-10
Vasunilashorn SM et al, 2019 [52]	USA	Case-Control Study	CAM	IL-6, IL-2, CRP, SERPINA3, HPX, ORM1, AZGP1
Nübel J et al, 2023 [53]	Germany	Cohort Study	CAM-ICU	NSE
Dillon St et al, 2023 [54]	USA	Case-control Study	CAM	ACAN, CFL1, CXCL11, H2AFZ, MUC1, NAMPT, INS, CD97, ICOS, PARK7, FAM107B, CD38, NGF, PPIF, THPO, DCN, MICA, HAPLN1, CTSV, GNLY, CXCL6, CCL2, MMP14, MSN, CHRDL1, PROC, CCL28, CTSD, FSTL1, IGFBP2, PTPN6, PRKCA
Zhang Y et al, 2023 [55]	China	Longitudinal Study	CAM	IL-6, sIL-6R
Liang F et al, 2023 [56]	USA	Cohort Study	CAM, MDAS	Tau-PT181, Tau-PT217
Leung JM et al, 2023 [57]	USA	Case-Control Study	CAM	NFL
Van Munster BC et al, 2010 [58]	Netherlands	Cohort Study	CAM	Cortisol, IL-6, IL-8, S100B
Peters van Ton AM et al, 2020 [59]	Netherlands	Case-Control Study	DSM-IV	NBL1, THY1, NrCAM, NCAN, TNKRSP21, DINER, CADM3, RGMA, CTSS, IL-1a, RSP01, N-Cdase, WFIKKN1, HAGH, CD200R1, IL-5ra, ENRAGE, MCP4, CRTAM, NEP, CST5, BCAN, CASP8, EFNA4, SCF, EZR, CX3CL1, HGF, TGF-B1, SMO2, CXCL6
Vasunilashorn SM et al, 2022 [60]	USA	Case-Control Study	CAM	CHI3L1, PF4, MICA, ADCYAP1, RETN, CD300C, CD274, FCGR3B, PAPP, TNFRSF1A, PLA2G2A, IL-6, TIMP1, THBS1, CD177, CKM, NAAA, ANP32B, KLKB1, BMP1, C4A, CCL27, CTSV, TNFSF9, CCL11, IL-25, AMN, STX1A, CCL16, CHKB, SERPING1, MIA, CCL27, CDH1, PLG, LRIG3

(Continued)

Table 1. (Continued)

Authors & year	Country	Type of Study	Delirium Assessment Methods	All Associated Proteins/Genes
Kaźmierski J et al, 2021 [61]	Poland	Cohort Study	CAM	MCP-1, CRP
Ye C et al, 2020 [62]	China	Cohort Study	CAM-ICU	IL-6, CHI3L1, S100B, Lp-PLA2, MIF, ICAM-1, VCAM-1, BACE1, a-SYN
Ritchie CW et al, 2014 [63]	England	Cross-Sectional Study	CAM	CRP
Plaschke K et al, 2010 [64]	Germany	Cohort Study	CAM-ICU	IL-6, Cortisol
Szwed K et al, 2021 [65]	Poland	Case-Control Study	CAM-ICU	NSP, GFAP
Erikson K et al, 2019 [66]	Finland	Cohort Study	CAM-ICU	S100B, IL-6
Yuan Y et al, 2020 [67]	China	Case-Control Study	CAM	IL-1b, IL-6, a-SYN
Khan SH et al, 2022 [68]	USA	Randomized Control Trial	CAM-ICU	CRP, IL-8, IL-10
Dönmezler S et al, 2023 [69]	Turkey	Case-Control Study	DSM-5	eGFR, proBNP
Wiredu K et al, 2023 [70]	USA	Case-Control Study	CAM	CRP, CH3L1, AACT, TIMP1, FGL1, SAA1, SAA2, IBP2, CAH3, CATB, PEPA3, ACTN1
Su LJ et al, 2023 [71]	China	Cohort Study	CAM-ICU	sTNFR-1, sTNFR-2
Shyam R et al, 2023a [72]	India	Case-Control Study	CAM-ICU	S100B
Tsui A et al, 2023 [73]	England	Cohort Study	DSM-IV	CRP
Menzenbach J et al, 2021 [74]	Germany	Cohort Study	CAM-ICU	CCL2, SDC-1
Boogaard MVD et al, 2011 [75]	Netherlands	Cross-Sectional Study	CAM-ICU	IL-8, MCP-1, PCT, Cortisol, TNF-a, MIF, IL-8, IL-1b, IL-1ra, IL-10
Hall RJ et al, 2013 [76]	Scotland	Cohort Study	CAM	S100B
Xu WB et al, 2019 [77]	China	Case-Control Study	CAM	sFGL2
Chai LV et al, 2021 [78]	China	Cohort Study	CAM	IL-6
Khan BA et al, 2013 [79]	USA	Cohort Study	CAM-ICU	S100B
Mao M et al, 2022 [80]	China	Cohort Study	CAM	PGE2, NFL, S100B, GFAP
Khan BA et al, 2020 [81]	USA	Cohort Study	CAM-ICU	IL-6, IL-8, IL-10, TNF-a, CRP, S100B
Neerland et al, 2016 [82]	Norway	Cohort Study	CAM	CRP, sIL-6R
Girard TD et al, 2012 [83]	USA	Cohort Study	CAM-ICU	MMP-9, Protein C, sTNFR1
Maes M et al, 2022 [84]	Thailand	Cohort Study	DRSR-98	GlutaR, AQP4, HSP60
Pfister D et al, 2008 [85]	USA	Cohort Study	CAM-ICU	CRP, S100B, Cortisol
Cerejeira J et al, 2012 [86]	Portugal	Cohort Study	CAM	CRP, IL-6, IL-8, IL-10
Plaschke K et al, 2023 [87]	Germany	Cohort Study	ICDSC, CAM-ICU	PON1, IgHV3, C1QC, THBS1, FGG
Wu X et al, 2023b [88]	China	Cohort Study	CAM, MDAS	Aβ42, P-tau, T-tau
Kim HJ et al, 2023 [89]	South Korean	Cohort Study	DSM-5	CRP
Rooij SE et al, 2007 [90]	Netherlands	Cohort Study	CAM	IL-6, IL-8
Wang B et al, 2022 [91]	China	Case-Control Study	CAM	APP
McNeil JB et al, 2019 [92]	USA	Cohort Study	CAM-ICU	PAI-1, IL-6
Klimiec Moskal et al, 2021 [93]	Poland	Cohort Study	CAM-ICU	Gal-3BP
Cape E et al, 2014 [94]	Netherlands	Cohort Study	CAM	IL-1b, IL-1ra
Lindblom RPF et al, 2018 [95]	Sweden	Cohort Study	CAM-ICU	TR4, EZH2, CHI3L1, IL-6, SFRP2, PMP2, RTN4R, GFAP, CX3CL1, ICAM-1
Skrede et al, 2015 [96]	Norway	Cohort Study	CAM	MCP-1
Brattinga B et al, 2022 [97]	Netherlands	Cohort Study	DOS scale	IL-6, IL-10, NGAL
Chen BY et al, 2019 [98]	China	Cohort Study	CAM-ICU	IL-6
Shen H et al, 2016 [99]	China	Cohort Study	CAM, DRS-R98	IGF-1
Shyam R et al, 2023b [100]	India	Case-Control Study	CAM-ICU	CRP

(Continued)

Table 1. (Continued)

Authors & year	Country	Type of Study	Delirium Assessment Methods	All Associated Proteins/Genes
Brown et al, 2023 [101]	USA	Cohort Study	CAM, CAM-ICU, DSM-5	NFL
Klimiec-Moskal et al, 2023 [102]	Poland	Cohort Study	bCAM, CAM-ICU	CRP
Imai T et al, 2023 [103]	Japan	Cohort Study	CAM	IL-6
Khan SH et al, 2023 [104]	USA	Case-Control Study	CAM-ICU, RASS	F9, MAN1A1, AGT, CP, C2, ITIH3

<https://doi.org/10.1371/journal.pone.0309827.t001>

delirium. They are Interlukitin-6 (IL-6), C-reactive protein (CRP), Interlukitin-8 (IL-8), S100B calcium-binding protein, Interlukitin-10 (IL-10), Tumor necrosis factor- α (TNF- α), Interlukitin-1b (IL-1b), Cortisol, Monocyte chemoattractant protein 1 (MCP-1), Glial fibrillary acidic protein (GFAP), Insulin-like-growth-factor-1 (IGF-1), Interlukitin-1 receptor antagonist (IL-1ra), and Neurofilament light polypeptide (NFL). Among them, the IL-6 was mostly reported (n = 29 studies), followed by the CRP (n = 16 studies); IL-8 (n = 11 studies); S100B (n = 10 studies); IL-10 (n = 8 studies); TNF- α (n = 7 studies); IL-1b (n = 6 studies); Cortisol (n = 5 studies); MCP-1 (n = 5 studies); and other proteins subsequently presented in Fig 3.

Based on the distribution of the proteins, we found the top 13 key proteins that were reported in a minimum of four studies. Table 2 describes the basic functionality and clinical justification of the most reported proteins. From the distribution of the functionality of the reported proteins, it has been clear that most of them are associated with cytokines and inflammatory functionality in the human body. Some of them functioned as neurotrophic factors

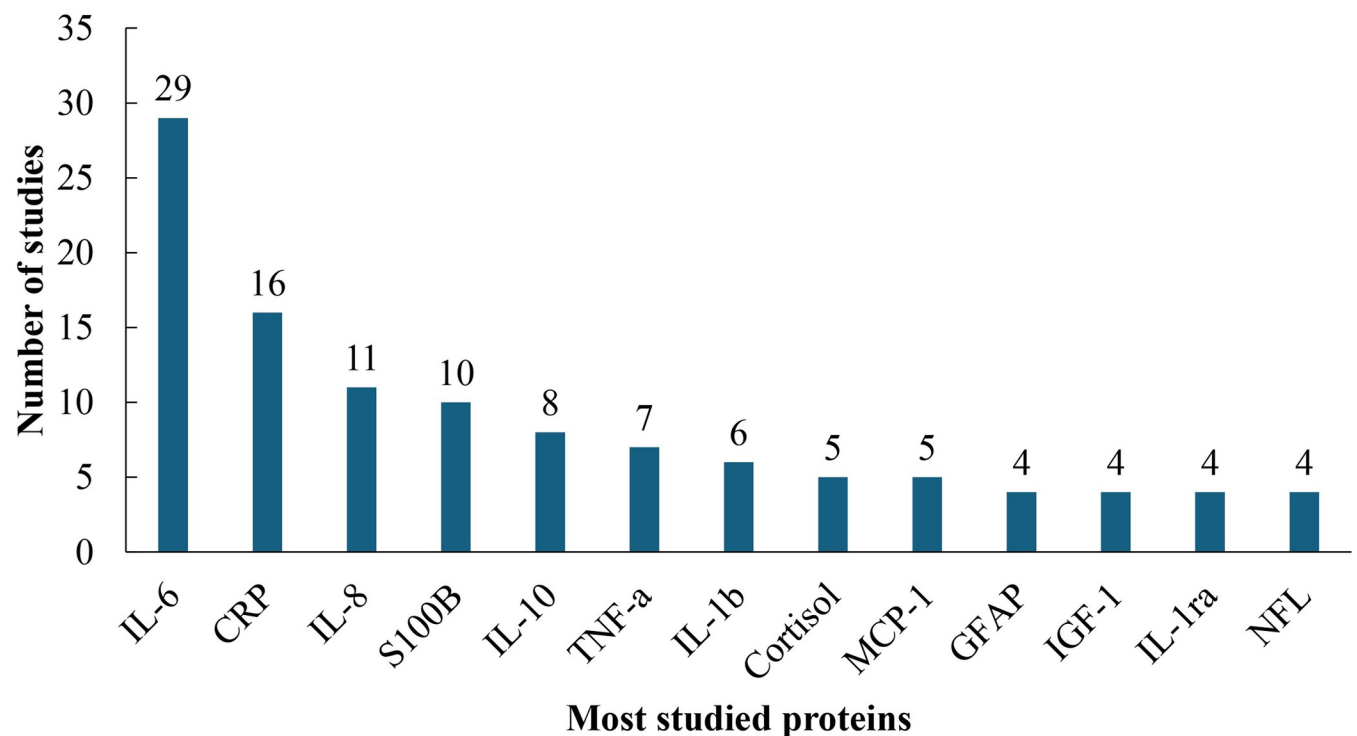


Fig 3. The distribution of the most-studied proteins collected from the included studies.

<https://doi.org/10.1371/journal.pone.0309827.g003>

Table 2. Properties of the 14 most studied proteins.

Annotated Proteins	Full description	Functionality	Clinical justifications
IL-6	Interlukitin-6	Cytokine	Inflammatory marker
CRP	C-reactive protein	Cytokine and Inflammation	Inflammatory marker
IL-8	Interlukitin-8	Cytokine	Inflammatory marker
S100 β	S100B, calcium binding protein	Cytokine and Neurotrophic factor	Tumor marker, brain damage marker
IL-10	Interlukitin-10	Cytokine	Inflammatory marker
TNF- α	Tumor necrosis factor-alpha	Cytokine	Inflammatory marker
IL-1B	Interlukitin-1b	Cytokine	Inflammatory marker
Cortisol	Cortisol	Hormone	Adrenal function indicator
MCP-1	Monocyte chemoattractant protein 1	Cytokine/Chemokine	Inflammation
GFAP	Glial fibrillary acidic protein	Intermediate filaments	Roles in neurodegenerative disorder, cell migration, mitosis, development
IGF-1	Insulin-like-growth-factor-1	Growth factor	Growth hormone deficiency diagnosis, pituitary function indicator
IL-1ra	Interleukin-1 receptor antagonist	Cytokine and Inflammation	Immune and inflammatory responses
NFL	Neurofilament light polypeptide	Filament proteins	Roles in neurodegenerative disorder, interconnection between axons and dendrites

<https://doi.org/10.1371/journal.pone.0309827.t002>

and growth factors. Based on their clinical justification and linkages, they are implicated in a variety of functional pathways, including those for are involved in different functional pathways like the inflammatory response, immune response, neurodegenerative disorder, growth, and brain damage functions, and many more.

Discussion

The review showed that most of the significantly associated proteins with delirium belong to the cytokines and inflammatory functionality groups. Although delirium is a multifactorial condition encompassing older age, alcohol or drug usage, severe comorbidities, and anesthesia. A study conducted by Liu et al 2018 [105] showed the cytokines and inflammatory proteins also revealed a strong association of a few of them with delirium.

The included studies reported one or more delirium-associated biomarkers proteins/genes however, there is not enough evidence to justify the use of one specific diagnostic or biomolecule as the sole risk or disease biomarker for delirium. This may happen due to methodological variations, distinct analytical procedures, a variety of patient populations, diagnostic criteria for identifying delirium, and the presence of complicating comorbidities. It was also difficult to determine the strength of the identified relationships between the genes/proteins and delirium.

Role of cytokines and inflammation in delirium

The included studies revealed a total of 189 unique gene-encoded proteins where some of the proteins occurred frequently such as the proinflammatory cytokines IL-6, IL-8, CRP, S100B, TNF- α , IL-1b, MCP-1, and IL-1ra; the anti-inflammatory cytokine IL-10. The studies suggest that the pro-inflammatory cytokines (PICs), including IL-6 and TNF- α , and anti-inflammatory cytokines, including IL-10, are significant subgroups of inflammatory processes and are crucial in the development of pain sensitivity [51, 106–108]. Since our included studies reported a significant relationship (either positive or negative) between cytokines and delirium

development, these indicate that there exists a huge interconnection between cytokines, inflammation, and delirium. The onset of delirium is caused by an early sign of systemic inflammation which indicates the involvement of the cytokine proteins [35, 41, 109–111]. Particularly, regarding its potential to serve as a delirium marker, IL-6 is one of the cytokines that has been examined the most among the included studies. The other cytokines namely, IL-8, MCP-1, IL-1ra, and the anti-inflammatory IL-10 show a significant stimulation of the major immune response pathways as well as in monocyte chemoattraction [29, 94, 96, 112–114]. Plasma IL-6 levels are correlated with delirium intensity and length in critically ill individuals, indicating that systemic inflammation plays a role in the onset and progression of delirium [62, 92]. Our studies suggested that the expression level of cytokine proteins changed significantly in delirious and non-delirious patients or preoperative and post-operative stages. In this aspect, the altered cytokine patterns point to an immune reaction that includes B-cell and T-cell stimulation, immunoglobulin production, and concurrent initiation of anti-inflammatory processes [29, 115, 116].

The CRP plays a novel role in the pathophysiology of delirium as it is associated with stress response, and inflammation and has a role in neurotransmitter activities [82, 117]. Among the investigated studies CRP was reported in 10 individual studies which indicate its significance. Ritchie et al, 2014 reported that CRP has a noticeable association with delirious patients having “musculoskeletal” problems [63]. Although there exists some inconsistency about the role of CRP in delirium as a potential biomarker, it can be focused on future in-depth research to clarify its role broadly.

As a pro-inflammatory cytokine, IL-1b is highly associated with delirium development as well as having a part in the etiology of early delirium [94]. It also plays a vital role in cholinergic activity, a route believed to be responsible for the pathophysiology of delirium [118]. Having some contradictory findings, IL-1b could not be served as a potential individual biomarker for delirium [111]. The S100B is considered as a calcium-binding protein that has involvement in astrocytes with the central nervous system (CNS) and is associated with delirium [66, 119, 120]. The presence of S100B in cerebrospinal fluid (CSF) indicates the early symptoms of Alzheimer’s dementia [75] which is one of the crucial adverse events for delirious patients. A pleiotropic cytokine TNF-a, is associated with several functional pathways including inflammation, necrosis, strong association with cognitive deterioration, apoptosis, and delirium as well [35, 121, 122]. Due to the strong association with cognitive decline (like Alzheimer’s disease), it’s difficult to announce TNF-a as a potential biomarker of delirium [111] which demands further research to clarify the specific role of TNF-a in delirium development.

Other proteins are also found as top studies proteins in our review namely, IGF-1, GFAP, and NFL. All of these are associated with delirium development in preoperative or postoperative stages. IGF-1 is known as a neuroprotective and growth factor which involves in neurogenesis and also may inhibit cytotoxic cytokines, leads pro-inflammation [123, 124]. In our review, three studies reported a negative association of IGF-1 with delirium [28, 91, 124] and one study reported a positive relationship [31]. Due to the linkage with the pathophysiology of Alzheimer’s disease [125, 126], it is still considered an inconsistency among biomarkers of delirium. The study revealed that the likelihood of delirium recovery may be influenced by lower levels of IGF-1 and the lack of the APOE-e4 genotype among female patients [127]. Three studies identify the increased level of GFAP proteins which is significantly associated with delirium [65, 80, 95] in our review. The NFL are associated with ongoing axonal complications and considered as a novel biomarker of Alzheimer’s disease [128, 129], variety of neurological disorders [130, 131].

The above-mentioned discussion indicates the importance of reported significant biomolecules for delirium development. Several cytokines and inflammatory proteins are highly

reported mentioning the association with delirium. Future research and deeper molecular investigation should focus on cytokines and inflammation related proteins and their associated signaling pathways to decipher the pathophysiology of delirium.

Implications

This review concludes that cytokines and inflammatory proteins play a crucial role in delirium development. Delirium's pathophysiology is multifactorial, and diverse sampling types should be considered for molecular studies [111]. Further studies involving the gene expression study could be a reliable source to identify the differentially expressed genes and proteins associated with delirium. The gene expression data analysis may assist to clarify the pathogenesis of delirium as well as the functional pathways. Future in-depth research on epigenetic analysis and genome-wide association studies may also benefit to identify potential biomarkers which will eventually help in delirium diagnosis and therapeutics. In this regard, this study will be a basis for further proteomic research in delirium.

Study limitations

This review focused on delirium in humans whether they were identified in ICU or any hospital settings. Studies including Alzheimer's disease and dementia have not been included to keep the study rigorous and focused solely on delirium. This review's search for relevant studies may have been limited by the databases used, which might result in the disappearance of potential studies. The current study covered the timeframe between January 2000 and December 2023, therefore, studies before 2000 and after 2023 have not been included. Besides that, this study retrieved published articles from PubMed, Scopus, and EBSCOhost (CINAHL, Medline) databases. If there exist any potential published studies outside of these databases, the article might be missing from this study. The current study focused on accumulating the proteomic biomarkers only, therefore considering the proteins and the gene-encoded proteins to investigate. No specific diagnostic and prognostic proteomic biomarker could be identified through this study. The reported relationship between the proteins and delirium was only considered when the direction of the relationship (positive or negative) was ignored. Therefore, the properties of upregulation or downregulation of proteins could not be described in this review which demands further studies to investigate the differentially expressed genes/proteins identification. Moreover, the confounding variables, estimation process variation, lack of random allocation, study setting were not considered and completely ruled out.

Conclusion

This study magnifies the significant information regarding delirium-associated proteomic biomarkers. We have summarized the 13 most studied proteins (IL-6, CRP, IL-8, S100B, IL-10, TNF- α , IL-1 β , Cortisol, MCP-1, GFAP, IGF-1, IL-1 α and NFL) about delirium. Notably, cytokine and inflammatory proteomic factors are the most crucial influencer for delirium development and the ultimate stage of delirium, found in this study. Inconsistency among the proteomic biomarkers and the lack of knowledge about the entire pathophysiological process of delirium demand more in-depth molecular studies to decipher the core knowledge of the molecular functionality of delirium. More studies need to be conducted to identify the exclusive causal genomic and proteomic biomarker of delirium which can be investigated as prognostic, diagnostic, and therapeutic target biomolecules. The summarization of the current information on delirium-associated proteins that has been done in this study might serve as a guide for further research and in-depth investigation of delirium.

Supporting information

S1 File. The search sentences are used in different databases and outcomes.
(PDF)

S2 File. The overall quality appraisal scores.
(XLSX)

S3 File. The PRISMA checklist.
(DOCX)

S4 File. Delirium-associated 189 gene-encoded unique proteins.
(XLSX)

S1 Table. The patient's demographic and study characteristics table.
(DOCX)

S2 Table. The screened studies with decision, entire data matrix.
(XLSX)

Acknowledgments

The authors are acknowledging the reviewer's contribution to assist in the study screening and selection process, their comments, and suggestions for improving the quality of the manuscript.

Author Contributions

Conceptualization: Md Parvez Mosharaf.

Data curation: Md Parvez Mosharaf, Rashidul Alam Mahumud.

Formal analysis: Md Parvez Mosharaf.

Methodology: Md Parvez Mosharaf, Rashidul Alam Mahumud.

Supervision: Khorshed Alam, Jeff Gow, Rashidul Alam Mahumud.

Writing – original draft: Md Parvez Mosharaf.

Writing – review & editing: Md Parvez Mosharaf, Khorshed Alam, Jeff Gow, Rashidul Alam Mahumud.

References

1. Francis J, Kapoor WN. Delirium in hospitalized elderly. *J Gen Intern Med.* 1990; 5: 65–79. <https://doi.org/10.1007/BF02602312> PMID: 2405116
2. American Psychiatric Association (APA). *Diagnostic and Statistical Manual of Mental Disorders (DSM-5-TR)*. Am Psychiatr Assoc. 2023. <https://www.psychiatry.org/psychiatrists/practice/dsm>
3. AGS/NIA Delirium Conference Writing Group PC and F. The American Geriatrics Society/National Institute on Aging Bedside-to-Bench Conference: Research Agenda on Delirium in Older Adults. *J Am Geriatr Soc.* 2015; 63: 843–52. <https://doi.org/10.1111/jgs.13406> PMID: 25834932
4. Marcantonio ER. Delirium in Hospitalized Older Adults. Solomon CG, editor. *N Engl J Med.* 2017; 377: 1456–1466. <https://doi.org/10.1056/NEJMcp1605501> PMID: 29020579
5. Poulsen LM, Estrup S, Mortensen CB, Andersen-Ranberg NC. Delirium in Intensive Care. *Curr Anesthesiol Rep.* 2021; 11: 516–523. <https://doi.org/10.1007/s40140-021-00476-z> PMID: 34493931
6. Inouye SK, Westendorp RGJ, Saczynski JS. Delirium in elderly people. *The Lancet.* 2014. pp. 911–922. [https://doi.org/10.1016/S0140-6736\(13\)60688-1](https://doi.org/10.1016/S0140-6736(13)60688-1) PMID: 23992774

7. Marcantonio ER. Delirium. *Ann Intern Med.* 2011; 154: ITC6. <https://doi.org/10.7326/0003-4819-154-11-201106070-01006> PMID: 21646553
8. Yang FM, Marcantonio ER, Inouye SK, Kiely DK, Rudolph JL, Fearing MA, et al. Phenomenological subtypes of delirium in older persons: Patterns, prevalence, and prognosis. *Psychosomatics.* 2009; 50: 248–254. <https://doi.org/10.1176/appi.psy.50.3.248> PMID: 19567764
9. Gleason LJ, Schmitt EM, Kosar CM, Tabloski P, Saczynski JS, Robinson T, et al. Effect of delirium and other major complications on outcomes after elective surgery in older adults. *JAMA Surg.* 2015; 150: 1134–1140. <https://doi.org/10.1001/jamasurg.2015.2606> PMID: 26352694
10. Maldonado JR. Neuropathogenesis of delirium: Review of current etiologic theories and common pathways. *Am J Geriatr Psychiatry.* 2013; 21: 1190–1222. <https://doi.org/10.1016/j.jagp.2013.09.005> PMID: 24206937
11. Marcantonio ER, Rudolph JL, Culley D, Crosby G, Alsop D, Inouye SK. Serum biomarkers for delirium. *J Gerontol A Biol Sci Med Sci.* 2006; 61: 1281–6. <https://doi.org/10.1093/gerona/61.12.1281> PMID: 17234821
12. Androsova G, Krause R, Winterer G, Schneider R. Biomarkers of postoperative delirium and cognitive dysfunction. *Front Aging Neurosci.* 2015; 7. <https://doi.org/10.3389/fnagi.2015.00112> PMID: 26106326
13. Hansen N, Krsiuk I, Titsch T. Neural autoantibodies in delirium. *J Autoimmun.* 2021; 125: 102740. <https://doi.org/10.1016/j.jaut.2021.102740> PMID: 34757245
14. Hall RJ, Watne LO, Cunningham E, Zetterberg H, Shenkin SD, Wyller TB, et al. CSF biomarkers in delirium: a systematic review. *Int J Geriatr Psychiatry.* 2018; 33: 1479–1500. <https://doi.org/10.1002/gps.4720> PMID: 28585290
15. Hsieh TT, Fong TG, Marcantonio ER, Inouye SK. Cholinergic deficiency hypothesis in delirium: a synthesis of current evidence. *J Gerontol A Biol Sci Med Sci.* 2008; 63: 764–72. <https://doi.org/10.1093/gerona/63.7.764> PMID: 18693233
16. Khan BA, Zawahiri M, Campbell NL, Boustani MA. Biomarkers for delirium—A review. *Journal of the American Geriatrics Society.* NIH Public Access; 2011. p. S256. <https://doi.org/10.1111/j.1532-5415.2011.03702.x> PMID: 22091570
17. Elie M, Cole MG, Primeau FJ, Bellavance F. Delirium risk factors in elderly hospitalized patients. *Journal of General Internal Medicine.* 1998. pp. 204–212. <https://doi.org/10.1046/j.1525-1497.1998.00047.x> PMID: 9541379
18. Mosharaf MP, Reza MS, Gov E, Mahumud RA, Mollah MNH. Disclosing Potential Key Genes, Therapeutic Targets and Agents for Non-Small Cell Lung Cancer: Evidence from Integrative Bioinformatics Analysis. *Vaccines.* 2022; 10: 771. <https://doi.org/10.3390/vaccines10050771> PMID: 35632527
19. Mosharaf MP, Reza MS, Kibria MK, Ahmed FF, Kabir MH, Hasan S, et al. Computational identification of host genomic biomarkers highlighting their functions, pathways and regulators that influence SARS-CoV-2 infections and drug repurposing. *Sci Rep.* 2022; 12: 4279. <https://doi.org/10.1038/s41598-022-08073-8> PMID: 35277538
20. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. *Int J Surg.* 2021; 88: 105906. <https://doi.org/10.1016/j.ijsu.2021.105906> PMID: 33789826
21. Moola S, Munn Z, Sears K, Sfetcu R, Currie M, Lisy K, et al. Conducting systematic reviews of association (etiology): The Joanna Briggs Institute’s approach. *Int J Evid Based Healthc.* 2015; 13: 163–169. <https://doi.org/10.1097/XEB.000000000000064> PMID: 26262566
22. Joanna Briggs Institute (JBI). JBI’s critical appraisal tools. *Fac Heal Med Sci Univ Adelaide SA 5006 Adelaide, Aust.* 2022; 2–6. <https://jbi.global/critical-appraisal-tools>
23. Mahumud RA, Kamara JK, Renzaho AMN. The epidemiological burden and overall distribution of chronic comorbidities in coronavirus disease-2019 among 202,005 infected patients: evidence from a systematic review and meta-analysis. *Infection.* 2020. pp. 813–833. <https://doi.org/10.1007/s15010-020-01502-8> PMID: 32813220
24. Porto De Toledo I, Stefani FM, Porporatti AL, Mezzomo LA, Peres MA, Flores-Mir C, et al. Prevalence of otologic signs and symptoms in adult patients with temporomandibular disorders: a systematic review and meta-analysis. *Clin Oral Investig.* 2017; 21: 597–605. <https://doi.org/10.1007/s00784-016-1926-9> PMID: 27511214
25. Munn Z, MClinSc SM, Lisy K, Riitano D, Tufanaru C. Methodological guidance for systematic reviews of observational epidemiological studies reporting prevalence and cumulative incidence data. *Int J Evid Based Healthc.* 2015; 13: 147–153. <https://doi.org/10.1097/XEB.000000000000054> PMID: 26317388

26. Mahumud RA, Ali MA, Kundu S, Rahman MA, Kamara JK, Renzaho AMN. Effectiveness of COVID-19 Vaccines against Delta Variant (B.1.617.2): A Meta-Analysis. *Vaccines*. 2022; 10: 277. <https://doi.org/10.3390/vaccines10020277> PMID: 35214737
27. Adamis D, Lunn M, Martin FC, Treloar A, Gregson N, Hamilton G, et al. Cytokines and IGF-I in delirious and non-delirious acutely ill older medical inpatients. *Age Ageing*. 2009; 38: 326–32; discussion 251. <https://doi.org/10.1093/ageing/afp014> PMID: 19269948
28. Egberts A, Wijnbeld EHA, Fekkes D, van der Ploeg MA, Ziere G, Hooijkaas H, et al. Neopterin: a potential biomarker for delirium in elderly patients. *Dement Geriatr Cogn Disord*. 2015; 39: 116–24. <https://doi.org/10.1159/000366410> PMID: 25413160
29. Hirsch J, Vacas S, Terrando N, Yuan M, Sands LP, Kramer J, et al. Perioperative cerebrospinal fluid and plasma inflammatory markers after orthopedic surgery. *J Neuroinflammation*. 2016; 13: 211. <https://doi.org/10.1186/s12974-016-0681-9> PMID: 27577265
30. Kazmierski J, Banys A, Latek J, Bourke J, Jaszewski R. Raised IL-2 and TNF- α concentrations are associated with postoperative delirium in patients undergoing coronary-artery bypass graft surgery. *Int Psychogeriatrics*. 2014; 26: 845–855. <https://doi.org/10.1017/S1041610213002378> PMID: 24345656
31. Miao S, Shen P, Zhang Q, Wang H, Shen J, Wang G, et al. Neopterin and Mini-Mental State Examination scores, two independent risk factors for postoperative delirium in elderly patients with open abdominal surgery. *J Cancer Res Ther*. 2018; 14: 1234–1238. <https://doi.org/10.4103/0973-1482.192764> PMID: 30488836
32. Ritter C, Tomasi CD, Dal-Pizzol F, Pinto BB, Dyson A, de Miranda AS, et al. Inflammation biomarkers and delirium in critically ill patients. *Crit Care*. 2014; 18: R106. <https://doi.org/10.1186/cc13887> PMID: 24886875
33. Sun L, Jia P, Zhang J, Zhang X, Zhang Y, Jiang H, et al. Production of inflammatory cytokines, cortisol, and Aβ1–40 in elderly oral cancer patients with postoperative delirium. *Neuropsychiatr Dis Treat*. 2016; Volume 12: 2789–2795. <https://doi.org/10.2147/NDT.S113077> PMID: 27822051
34. Van Munster BC, Korevaar JC, Zwinderman AH, Levi M, Wiersinga WJ, De Rooij SE. Time-course of cytokines during delirium in elderly patients with hip fractures. *J Am Geriatr Soc*. 2008; 56: 1704–1709. <https://doi.org/10.1111/j.1532-5415.2008.01851.x> PMID: 18691278
35. Vasunilashorn SM, Ngo L, Inouye SK, Libermann TA, Jones RN, Alsop DC, et al. Cytokines and Postoperative Delirium in Older Patients Undergoing Major Elective Surgery. *Journals Gerontol—Ser A Biol Sci Med Sci*. 2015; 70: 1289–1295. <https://doi.org/10.1093/gerona/glv083> PMID: 26215633
36. Liu X, Wang Y, Wu J, Ye C, Ma D, Wang E. Emergence delirium and postoperative delirium associated with high plasma NfL and GFAP: an observational study. *Front Med*. 2023; 10. <https://doi.org/10.3389/fmed.2023.1107369> PMID: 37576000
37. Sun Y, Peng H-P, Wu T-T. Postoperative C-Reactive Protein Predicts Postoperative Delirium in Colorectal Cancer Following Surgery. *Clin Interv Aging*. 2023; 18: 559–570. <https://doi.org/10.2147/CIA.S387117> PMID: 37038607
38. Oren RL, Kim EJ, Leonard AK, Rosner B, Chibnik LB, Das S, et al. Age-dependent differences and similarities in the plasma proteomic signature of postoperative delirium. *Sci Rep*. 2023; 13: 7431. <https://doi.org/10.1038/s41598-023-34447-7> PMID: 37156856
39. Wu X, Zhang N, Zhou B, Liu S, Wang F, Wang J, et al. Alcohol consumption may be associated with postoperative delirium in the elderly: the PNDABLE study. *BMC Anesthesiol*. 2023; 23: 222. <https://doi.org/10.1186/s12871-023-02178-x> PMID: 37353780
40. Ruhnau J, Müller J, Nowak S, Strack S, Sperlich D, Pohl A, et al. Serum Biomarkers of a Pro-Neuroinflammatory State May Define the Pre-Operative Risk for Postoperative Delirium in Spine Surgery. *Int J Mol Sci*. 2023; 24: 10335. <https://doi.org/10.3390/ijms241210335> PMID: 37373482
41. Westhoff D, Witlox J, Koenderman L, Kalisvaart KJ, de Jonghe JFM, van Stijn MFM, et al. Preoperative cerebrospinal fluid cytokine levels and the risk of postoperative delirium in elderly hip fracture patients. *J Neuroinflammation*. 2013; 10: 122. <https://doi.org/10.1186/1742-2094-10-122> PMID: 24093540
42. Heinrich M, Sieg M, Kruppa J, Nürnberg P, Schreier PH, Heilmann-Heimbach S, et al. Association between genetic variants of the cholinergic system and postoperative delirium and cognitive dysfunction in elderly patients. *BMC Med Genomics*. 2021; 14: 248. <https://doi.org/10.1186/s12920-021-01071-1> PMID: 34674705
43. van Munster BC, Yazdanpanah M, Tanck MWT, de Rooij SEJA, van de Giessen E, Sijbrands EJG, et al. Genetic polymorphisms in the DRD2, DRD3, and SLC6A3 gene in elderly patients with delirium. *Am J Med Genet B Neuropsychiatr Genet*. 2010; 153B: 38–45. <https://doi.org/10.1002/ajmg.b.30943> PMID: 19309018
44. Terrelonge M, LaHue SC, Tang C, Movsesyan I, Pullinger CR, Dubal DB, et al. KIBRA, MTNR1B, and FKBP5 genotypes are associated with decreased odds of incident delirium in elderly post-surgical patients. *Sci Rep*. 2022; 12: 556. <https://doi.org/10.1038/s41598-021-04416-z> PMID: 35017578

45. Yamanashi T, Saito T, Yu T, Alario A, Comp K, Crutchley KJ, et al. DNA methylation in the TNF-alpha gene decreases along with aging among delirium inpatients. *Neurobiol Aging*. 2021; 105: 310–317. <https://doi.org/10.1016/j.neurobiolaging.2021.05.005> PMID: 34192631
46. Yamanashi T, Nagao T, Wahba NE, Marra PS, Crutchley KJ, Meyer AA, et al. DNA methylation in the inflammatory genes after neurosurgery and diagnostic ability of post-operative delirium. *Transl Psychiatry*. 2021; 11: 627. <https://doi.org/10.1038/s41398-021-01752-6> PMID: 34887385
47. Steimer M, Kaiser S, Ulbrich F, Kalbhenn J, Bürkle H, Schallner N. Expression of HO1 and PER2 can predict the incidence of delirium in trauma patients with concomitant brain injury. *Sci Rep*. 2021; 11: 15388. <https://doi.org/10.1038/s41598-021-94773-6> PMID: 34321570
48. Nekrosius D, Kaminskaite M, Jokubka R, Pranckeviciene A, Lideikis K, Tamasauskas A, et al. Association of COMT Val158Met Polymorphism With Delirium Risk and Outcomes After Traumatic Brain Injury. *J Neuropsychiatry Clin Neurosci*. 2019; 31: 298–305. <https://doi.org/10.1176/appi.neuropsych.18080195> PMID: 31046593
49. Rhee J, Kuznetsov A, McKay T, Lyons M, Houstis N, Mekkonen J, et al. Serum Proteomics of Older Patients Undergoing Major Cardiac Surgery: Identification of Biomarkers Associated With Postoperative Delirium. *Front Aging Neurosci*. 2021; 13: 1–11. <https://doi.org/10.3389/fnagi.2021.699763> PMID: 34456709
50. Ballweg T, White M, Parker M, Casey C, Bo A, Farahbakhsh Z, et al. Association between plasma tau and postoperative delirium incidence and severity: a prospective observational study. *Br J Anaesth*. 2021; 126: 458–466. <https://doi.org/10.1016/j.bja.2020.08.061> PMID: 33228978
51. Tang C, Hu Y, Zhang Z, Wei Z, Wang H, Geng Q, et al. Dexmedetomidine with sufentanil in intravenous patient-controlled analgesia for relief from postoperative pain, inflammation and delirium after esophageal cancer surgery. *Biosci Rep*. 2020; 40: 1–12. <https://doi.org/10.1042/BSR20193410> PMID: 32343308
52. Vasunilashorn SM, Ngo LH, Chan NY, Zhou W, Dillon ST, Otu HH, et al. Development of a Dynamic Multi-Protein Signature of Postoperative Delirium. *J Gerontol A Biol Sci Med Sci*. 2019; 74: 261–268. <https://doi.org/10.1093/gerona/gly036> PMID: 29529166
53. Nübel J, Buhre C, Hoffmeister M, Oess S, Labrenz O, Jost K, et al. Association between Neuron-Specific Enolase, Memory Function, and Postoperative Delirium after Transfemoral Aortic Valve Replacement. *J Cardiovasc Dev Dis*. 2023; 10: 441. <https://doi.org/10.3390/jcdd10110441> PMID: 37998499
54. Dillon ST, Vasunilashorn SM, Otu HH, Ngo L, Fong T, Gu X, et al. Aptamer-Based Proteomics Measuring Preoperative Cerebrospinal Fluid Protein Alterations Associated with Postoperative Delirium. *Biomolecules*. 2023; 13: 1395. <https://doi.org/10.3390/biom13091395> PMID: 37759795
55. Zhang Y, Hu J, Zuo W, He P, Xue Q, Feng X, et al. Longitudinal Profiling of Plasma Cytokines and Its Association With Postoperative Delirium in Elderly Patients Undergoing Major Lower Limb Surgery: A Prospective Observational Study. *Anesth Analg*. 2023; 136: 34–42. <https://doi.org/10.1213/ANE.0000000000006250> PMID: 36534715
56. Liang F, Baldyga K, Quan Q, Khatri A, Choi S, Wiener-Kronish J, et al. Preoperative Plasma Tau-PT217 and Tau-PT181 Are Associated with Postoperative Delirium. *Ann Surg*. 2023; 277: E1232–E1238. <https://doi.org/10.1097/SLA.0000000000005487> PMID: 35794069
57. Leung JM, Rojas JC, Tang C, Chan B, Lario-Lago A, Boxer AL, et al. Presence of Preoperative Neurodegeneration Biofluid Markers in Patients with Postoperative Delirium. *Anesthesiology*. 2023; 139: 432–443. <https://doi.org/10.1097/ALN.0000000000004666> PMID: 37364279
58. van Munster BC, Bisschop PH, Zwiderman AH, Korevaar JC, Endert E, Wiersinga WJ, et al. Cortisol, interleukins and S100B in delirium in the elderly. *Brain Cogn*. 2010; 74: 18–23. <https://doi.org/10.1016/j.bandc.2010.05.010> PMID: 20580479
59. Peters van Ton AM, Verbeek MM, Alkema W, Pickkers P, Abdo WF. Downregulation of synapse-associated protein expression and loss of homeostatic microglial control in cerebrospinal fluid of infectious patients with delirium and patients with Alzheimer's disease. *Brain Behav Immun*. 2020; 89: 656–667. <https://doi.org/10.1016/j.bbi.2020.06.027> PMID: 32592865
60. Vasunilashorn SM, Dillon ST, Chan NY, Fong TG, Joseph M, Tripp B, et al. Proteome-Wide Analysis Using SOMAscan Identifies and Validates Chitinase-3-Like Protein 1 as a Risk and Disease Marker of Delirium Among Older Adults Undergoing Major Elective Surgery. Newman AB, editor. *J Gerontol A Biol Sci Med Sci*. 2022; 77: 484–493. <https://doi.org/10.1093/gerona/glaa326> PMID: 35239952
61. Kaźmierski J, Miler P, Pawlak A, Jerczyńska H, Woźniak J, Frankowska E, et al. Elevated Monocyte Chemoattractant Protein-1 as the Independent Risk Factor of Delirium after Cardiac Surgery. A Prospective Cohort Study. *J Clin Med*. 2021; 10: 1587. <https://doi.org/10.3390/jcm10081587> PMID: 33918634
62. Ye C, Zhang Y, Luo S, Cao Y, Gao F, Wang E. Correlation of Serum BACE1 With Emergence Delirium in Postoperative Patients: A Preliminary Study. *Front Aging Neurosci*. 2020; 12: 1–7. <https://doi.org/10.3389/fnagi.2020.555594> PMID: 33192455

63. Ritchie CW, Newman TH, Leurent B, Sampson EL. The association between C-reactive protein and delirium in 710 acute elderly hospital admissions. *Int psychogeriatrics*. 2014; 26: 717–24. <https://doi.org/10.1017/S1041610213002433> PMID: 24460925
64. Plaschke K, Fichtenkamm P, Schramm C, Hauth S, Martin E, Verch M, et al. Early postoperative delirium after open-heart cardiac surgery is associated with decreased bispectral EEG and increased cortisol and interleukin-6. *Intensive Care Med*. 2010; 36: 2081–9. <https://doi.org/10.1007/s00134-010-2004-4> PMID: 20689917
65. Szwed K, Szwed M, Kozakiewicz M, Karlowska-Pik J, Soja-Kukiela N, Bartoszevska A, et al. Circulating MicroRNAs and Novel Proteins as Potential Biomarkers of Neurological Complications after Heart Bypass Surgery. *J Clin Med*. 2021; 10: 3091. <https://doi.org/10.3390/jcm10143091> PMID: 34300256
66. Erikson K, Ala-Kokko TI, Koskenkari J, Liisanantti JH, Kamakura R, Herzig KH, et al. Elevated serum S-100 β in patients with septic shock is associated with delirium. *Acta Anaesthesiol Scand*. 2019; 63: 69–73. <https://doi.org/10.1111/aas.13228> PMID: 30079511
67. Yuan Y, Li Z, Yang N, Han Y, Ji X, Han D, et al. Exosome α -Synuclein Release in Plasma May be Associated With Postoperative Delirium in Hip Fracture Patients. *Front Aging Neurosci*. 2020; 12: 1–10. <https://doi.org/10.3389/fnagi.2020.00067> PMID: 32231560
68. Khan SH, Lindroth H, Jawed Y, Wang S, Nasser J, Seyffert S, et al. Serum Biomarkers in Postoperative Delirium After Esophagectomy. *Ann Thorac Surg*. 2022; 113: 1000–1007. <https://doi.org/10.1016/j.athoracsur.2021.03.035> PMID: 33774004
69. Dönmezler S, Uysal A, Kurt İ, Özmen D, Güçlü O. Common Biomarkers Associated With Delirium in Hospitalised Patients With COVID-19 at the Epicentre of Turkish Coronavirus Outbreak: A Case Control Study. *Arch Neuropsychiatry*. 2022; 60: 17–22. <https://doi.org/10.29399/npa.28128> PMID: 36911570
70. Wiredu K, O'Connor S, Naseem H, Brauer BL, Kettenbach AN, Frost HR, et al. Intraoperative plasma proteomic changes in cardiac surgery: In search of biomarkers of post-operative delirium. *Proteomics Clin Appl*. 2023; 17: e2200066. <https://doi.org/10.1002/prca.202200066> PMID: 36567636
71. Su L-J, Chen M-J, Yang R, Zou H, Chen T-T, Li S-L, et al. Plasma biomarkers and delirium in critically ill patients after cardiac surgery: A prospective observational cohort study. *Heart Lung*. 2023; 59: 139–145. <https://doi.org/10.1016/j.hrtlng.2023.02.010> PMID: 36801548
72. Shyam R, Solanki M, Patel M, Sachan R, Ali W. S100B as a predictor of delirium in critically ill obstetric patients: A nested case—control study. *Int J Crit Illn Inj Sci*. 2023; 13: 125. https://doi.org/10.4103/ijciis.ijciis_19_23 PMID: 38023577
73. Tsui A, Yeo N, Searle SD, Bowden H, Hoffmann K, Hornby J, et al. Extremes of baseline cognitive function determine the severity of delirium: a population study. *Brain*. 2023; 146: 2132–2141. <https://doi.org/10.1093/brain/awad062> PMID: 36856697
74. Menzenbach J, Frede S, Petras J, Guttenthaler V, Kirfel A, Neumann C, et al. Perioperative vascular biomarker profiling in elective surgery patients developing postoperative delirium: A prospective cohort study. *Biomedicines*. 2021; 9: 553. <https://doi.org/10.3390/biomedicines9050553> PMID: 34063403
75. van den Boogaard M, Kox M, Quinn KL, van Achterberg T, van der Hoeven JG, Schoonhoven L, et al. Biomarkers associated with delirium in critically ill patients and their relation with long-term subjective cognitive dysfunction; indications for different pathways governing delirium in inflamed and noninflamed patients. *Crit Care*. 2011; 15: R297. <https://doi.org/10.1186/cc10598> PMID: 22206727
76. Hall RJ, Ferguson KJ, Andrews M, Green AJE, White TO, Armstrong IR, et al. Delirium and cerebrospinal fluid S100B in hip fracture patients: A preliminary study. *Am J Geriatr Psychiatry*. 2013; 21: 1239–1243. <https://doi.org/10.1016/j.jagp.2012.12.024> PMID: 23602305
77. Xu W Bin, Hu QH, Wu CN, Fan ZK, Song ZF. Serum soluble fibrinogen-like protein 2 concentration predicts delirium after acute pancreatitis. *Brain Behav*. 2019; 9: e01261. <https://doi.org/10.1002/brb3.1261> PMID: 30884164
78. Lv XC, Lin Y, Wu Q song, Wang L, Hou Y ting, Dong Y, et al. Plasma interleukin-6 is a potential predictive biomarker for postoperative delirium among acute type a aortic dissection patients treated with open surgical repair. *J Cardiothorac Surg*. 2021; 16: 146. <https://doi.org/10.1186/s13019-021-01529-4> PMID: 34044881
79. Khan BA, Farber MO, Campbell N, Perkins A, Prasad NK, Hui SL, et al. S100 calcium binding protein B as a biomarker of delirium duration in the intensive care unit—An exploratory analysis. *Int J Gen Med*. 2013; 6: 855–861. <https://doi.org/10.2147/IJGM.S51004> PMID: 24324346
80. Mao M, Wang L yuan, Zhu L yue, Wang F, Ding Y, Tong J hua, et al. Higher serum PGE2 is a predictive biomarker for postoperative delirium following elective orthopedic surgery in elderly patients. *BMC Geriatr*. 2022; 22: 685. <https://doi.org/10.1186/s12877-022-03367-y> PMID: 35982410
81. Khan BA, Perkins AJ, Prasad NK, Shekhar A, Campbell NL, Gao S, et al. Biomarkers of Delirium Duration and Delirium Severity in the ICU. *Crit Care Med*. 2020; 48: 353–361. <https://doi.org/10.1097/CCM.0000000000004139> PMID: 31770149

82. Neerland BE, Hall RJ, Seljeflot I, Frihagen F, MacLulich AMJ, Raeder J, et al. Associations Between Delirium and Preoperative Cerebrospinal Fluid C-Reactive Protein, Interleukin-6, and Interleukin-6 Receptor in Individuals with Acute Hip Fracture. *J Am Geriatr Soc*. 2016; 64: 1456–63. <https://doi.org/10.1111/jgs.14238> PMID: 27341529
83. Girard TD, Ware LB, Bernard GR, Pandharipande PP, Thompson JL, Shintani AK, et al. Associations of markers of inflammation and coagulation with delirium during critical illness. *Intensive Care Med*. 2012; 38: 1965–1973. <https://doi.org/10.1007/s00134-012-2678-x> PMID: 22903241
84. Maes M, Thisayakorn P, Thipakorn Y, Tantavisut S, Sirivichayakul S, Vojdani A. Reactivity to neural tissue epitopes, aquaporin 4 and heat shock protein 60 is associated with activated immune—inflammatory pathways and the onset of delirium following hip fracture surgery. *Eur Geriatr Med*. 2022; 14: 99–112. <https://doi.org/10.1007/s41999-022-00729-y> PMID: 36520371
85. Pfister D, Siegemund M, Dell-Kuster S, Smielewski P, Rüegg S, Strebel SP, et al. Cerebral perfusion in sepsis-associated delirium. *Crit Care*. 2008; 12: R63. <https://doi.org/10.1186/cc6891> PMID: 18457586
86. Cerejeira JMS, Nogueira V, Luís P, Vaz-Serra A, Mukaetova-Ladinska EB. The cholinergic system and inflammation: Common pathways in delirium pathophysiology. *J Am Geriatr Soc*. 2012; 60: 669–675. <https://doi.org/10.1111/j.1532-5415.2011.03883.x> PMID: 22316182
87. Plaschke K, Brenner T, Fiedler MO, Hölle T, von der Forst M, Wolf RC, et al. Extracellular Vesicles as Possible Plasma Markers and Mediators in Patients with Sepsis-Associated Delirium—A Pilot Study. *Int J Mol Sci*. 2023; 24: 15781. <https://doi.org/10.3390/ijms242115781> PMID: 37958765
88. Wu X, Chi F, Wang B, Liu S, Wang F, Wang J, et al. Relationship between preoperative neutrophil-to-lymphocyte ratio and postoperative delirium: The PNDABLE and the PNDRFAP cohort studies. *Brain Behav*. 2023; 13: e3281. <https://doi.org/10.1002/brb3.3281> PMID: 37830267
89. Kim H-J, Lee S, Kim S-H, Lee S, Sim J-H, Ro Y-J. Association of C-reactive protein to albumin ratio with postoperative delirium and mortality in elderly patients undergoing hip fracture surgery: A retrospective cohort study in a single large center. *Exp Gerontol*. 2023; 172: 112068. <https://doi.org/10.1016/j.exger.2022.112068> PMID: 36549547
90. de Rooij SE, van Munster BC, Korevaar JC, Levi M. Cytokines and acute phase response in delirium. *J Psychosom Res*. 2007; 62: 521–525. <https://doi.org/10.1016/j.jpsychores.2006.11.013> PMID: 17467406
91. Wang B, Yin Z, Lin Y, Deng X, Liu F, Tao H, et al. Correlation between microRNA-320 and postoperative delirium in patients undergoing tibial fracture internal fixation surgery. *BMC Anesthesiol*. 2022; 22: 1–12. <https://doi.org/10.1186/s12871-022-01612-w> PMID: 35317728
92. McNeil JB, Hughes CG, Girard T, Ware LB, Ely EW, Chandrasekhar R, et al. Plasma biomarkers of inflammation, coagulation, and brain injury as predictors of delirium duration in older hospitalized patients. Mossello E, editor. *PLoS One*. 2019; 14: e0226412. <https://doi.org/10.1371/journal.pone.0226412> PMID: 31856187
93. Klimiec-Moskal E, Pasinska P, Kowalska K, Klimkowicz-Mrowiec A, Pera J, Slowik A, et al. Elevated plasma levels of galectin-3 binding protein are associated with post-stroke delirium—A pilot study. *J Neuroimmunol*. 2021; 356: 577579. <https://doi.org/10.1016/j.jneuroim.2021.577579> PMID: 33901789
94. Cape E, Hall RJ, van Munster BC, de Vries A, Howie SEM, Pearson A, et al. Cerebrospinal fluid markers of neuroinflammation in delirium: a role for interleukin-1 β in delirium after hip fracture. *J Psychosom Res*. 2014; 77: 219–25. <https://doi.org/10.1016/j.jpsychores.2014.06.014> PMID: 25124807
95. Lindblom RPF, Shen Q, Axén S, Landegren U, Kamali-Moghaddam M, Thelin S. Protein Profiling in Serum and Cerebrospinal Fluid Following Complex Surgery on the Thoracic Aorta Identifies Biological Markers of Neurologic Injury. *J Cardiovasc Transl Res*. 2018; 11: 503–516. <https://doi.org/10.1007/s12265-018-9835-8> PMID: 30367354
96. Skrede K, Wyller TB, Watne LO, Seljeflot I, Juliebø V. Is there a role for monocyte chemoattractant protein-1 in delirium? Novel observations in elderly hip fracture patients. *BMC Res Notes*. 2015; 8: 1–4. <https://doi.org/10.1186/s13104-015-1129-5> PMID: 25943983
97. Brattinga B, Plas M, Spikman JM, Rutgers A, De Haan JJ, Absalom AR, et al. The association between the inflammatory response following surgery and post-operative delirium in older oncological patients: A prospective cohort study. *Age Ageing*. 2022; 51: 1–9. <https://doi.org/10.1093/ageing/afab237> PMID: 35180288
98. Chen Y, Lu S, Wu Y, Shen Y, Zhao H, Ding S, et al. Change in Serum Level of Interleukin 6 and Delirium After Coronary Artery Bypass Graft. *Am J Crit Care*. 2019; 28: 462–470. <https://doi.org/10.4037/ajcc2019976> PMID: 31676521
99. Shen H, Shao Y, Chen J, Guo J. Insulin-Like Growth Factor-1, a Potential Predictive Biomarker for Postoperative Delirium Among Elderly Patients with Open Abdominal Surgery. *Curr Pharm Des*. 2016; 22: 5879–5883. <https://doi.org/10.2174/1381612822666160813234311> PMID: 27526790

100. Shyam R, Ali W, Patel ML, Solanki M, Sachan R. Correlation of C-reactive Protein with Delirium in Obstetrics Intensive Care Unit: A Tertiary Center Experience. *Indian J Crit Care Med.* 2023; 27: 315–321. <https://doi.org/10.5005/jp-journals-10071-24455> PMID: 37214122
101. Brown CH, Kim AS, Yanek L, Lewis A, Mandal K, Le L, et al. Association of perioperative plasma concentration of neurofilament light with delirium after cardiac surgery: a nested observational study. *Br J Anaesth.* 2023; 132: 312–319. <https://doi.org/10.1016/j.bja.2023.10.043> PMID: 38114355
102. Klimiec-Moskal E, Slowik A, Dziedzic T. Serum C-reactive protein adds predictive information for post-stroke delirium: The PROPOLIS study. *Acta Psychiatr Scand.* 2023; 147: 536–542. <https://doi.org/10.1111/acps.13489> PMID: 35996990
103. Imai T, Morita S, Hasegawa K, Goto T, Katori Y, Asada Y. Postoperative serum interleukin-6 level as a risk factor for development of hyperactive delirium with agitation after head and neck surgery with free tissue transfer reconstruction. *Auris Nasus Larynx.* 2023; 50: 777–782. <https://doi.org/10.1016/j.anl.2023.01.005> PMID: 36754686
104. Khan SH, Perkins AJ, Jawaid S, Wang S, Lindroth H, Schmitt RE, et al. Serum proteomic analysis in esophagectomy patients with postoperative delirium: A case-control study. *Heart Lung.* 2024; 63: 35–41. <https://doi.org/10.1016/j.hrtlng.2023.09.009> PMID: 37748302
105. Liu X, Yu Y, Zhu S. Inflammatory markers in postoperatedelirium (POD) and cognitive dysfunction (POCD): A meta-analysis of observational studies. *PLoS One.* 2018; 13. <https://doi.org/10.1371/journal.pone.0195659> PMID: 29641605
106. Tang C, Li J, Tai WL, Yao W, Zhao B, Hong J, et al. Sex differences in complex regional pain syndrome type I (CRPS-I) in mice. *J Pain Res.* 2017; Volume 10: 1811–1819. <https://doi.org/10.2147/JPR.S139365> PMID: 28831269
107. Sommer C, Kress M. Recent findings on how proinflammatory cytokines cause pain: peripheral mechanisms in inflammatory and neuropathic hyperalgesia. *Neurosci Lett.* 2004; 361: 184–7. <https://doi.org/10.1016/j.neulet.2003.12.007> PMID: 15135924
108. Grosu I, Lavand'homme P. Continuous regional anesthesia and inflammation: a new target. *Minerva Anesthesiol.* 2015; 81: 1001–9. Available: <http://www.ncbi.nlm.nih.gov/pubmed/25317576>
109. Liu P, Li Y, Wang X, Zou X, Zhang D, Wang D, et al. High serum interleukin-6 level is associated with increased risk of delirium in elderly patients after noncardiac surgery: a prospective cohort study. *Chin Med J (Engl).* 2013; 126: 3621–7. <https://doi.org/10.3760/CMA.J.ISSN.0366-6999.20130211> PMID: 24112153
110. Capri M, Yani SL, Chattat R, Fortuna D, Bucci L, Lanzarini C, et al. Preoperative, high IL-6 blood level is a risk factor of postoperative delirium onset in old patients. *Front Endocrinol (Lausanne).* 2014; 5: 173. <https://doi.org/10.3389/fendo.2014.00173> PMID: 25368603
111. Dunne SS, Coffey JC, Konje S, Gasior S, Clancy CC, Gulati G, et al. Biomarkers in delirium: A systematic review. *J Psychosom Res.* 2021; 147: 110530. <https://doi.org/10.1016/j.jpsychores.2021.110530> PMID: 34098376
112. Terrando N, Eriksson LI, Ryu JK, Yang T, Monaco C, Feldmann M, et al. Resolving postoperative neuroinflammation and cognitive decline. *Ann Neurol.* 2011; 70: 986–995. <https://doi.org/10.1002/ana.22664> PMID: 22190370
113. Vacas S, Degos V, Tracey KJ, Maze M. High-mobility group box 1 protein initiates postoperative cognitive decline by engaging bone marrow-derived macrophages. *Anesthesiology.* 2014; 120: 1160–7. <https://doi.org/10.1097/ALN.000000000000045> PMID: 24162463
114. Degos V, Maze M, Vacas S, Hirsch J, Guo Y, Shen F, et al. Bone fracture exacerbates murine ischemic cerebral injury. *Anesthesiology.* 2013; 118: 1362–1372. <https://doi.org/10.1097/ALN.0b013e31828c23f8> PMID: 23438676
115. Hauser CJ, Sursal T, Rodriguez EK, Appleton PT, Zhang Q, Itagaki K. Mitochondrial damage associated molecular patterns from femoral reamings activate neutrophils through formyl peptide receptors and P44/42 MAP kinase. *J Orthop Trauma.* 2010; 24: 534–538. <https://doi.org/10.1097/BOT.0b013e3181ec4991> PMID: 20736789
116. Zhang Q, Raouf M, Chen Y, Sumi Y, Sursal T, Junger W, et al. Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature.* 2010; 464: 104–7. <https://doi.org/10.1038/nature08780> PMID: 20203610
117. Toft K, Tontsch J, Abdelhamid S, Steiner L, Siegemund M, Hollinger A. Serum biomarkers of delirium in the elderly: a narrative review. *Ann Intensive Care.* 2019; 9: 76. <https://doi.org/10.1186/s13613-019-0548-1> PMID: 31263968
118. Terrando N, Monaco C, Ma D, Foxwell BMJ, Feldmann M, Maze M. Tumor necrosis factor-alpha triggers a cytokine cascade yielding postoperative cognitive decline. *Proc Natl Acad Sci U S A.* 2010; 107: 20518–22. <https://doi.org/10.1073/pnas.1014557107> PMID: 21041647

119. Avila-Funes JA, Ledesma-Heyer JP, Navarrete-Reyes AP, Chavira-Ramírez R, Boeck-Quirasco L, Aguilar-Navarro S. Association between high serum estradiol levels and delirium among hospitalized elderly women. *Rev Invest Clin*. 2015; 67: 20–24. Available: <http://www.ncbi.nlm.nih.gov/pubmed/25857580>
120. Pinto SS, Gottfried C, Mendez A, Gonçalves D, Karl J, Gonçalves CA, et al. Immunocontent and secretion of S100B in astrocyte cultures from different brain regions in relation to morphology. *FEBS Lett*. 2000; 486: 203–207. [https://doi.org/10.1016/s0014-5793\(00\)02301-2](https://doi.org/10.1016/s0014-5793(00)02301-2) PMID: 11119704
121. Kalliolias GD, Ivashkiv LB. TNF biology, pathogenic mechanisms and emerging therapeutic strategies. *Nat Rev Rheumatol*. 2016; 12: 49–62. <https://doi.org/10.1038/nrrheum.2015.169> PMID: 26656660
122. Holmes C, Cunningham C, Zotova E, Woolford J, Dean C, Kerr S, et al. Systemic inflammation and disease progression in alzheimer disease. *Neurology*. 2009; 73: 768–774. <https://doi.org/10.1212/WNL.0b013e3181b6bb95> PMID: 19738171
123. Jones JI, Clemmons DR. Insulin-like growth factors and their binding proteins: biological actions. *Endocr Rev*. 1995; 16: 3–34. <https://doi.org/10.1210/edrv-16-1-3> PMID: 7758431
124. Adamis D, Meagher D. Insulin-Like Growth Factor I and the Pathogenesis of Delirium: A Review of Current Evidence. *J Aging Res*. 2011; 2011: 1–11. <https://doi.org/10.4061/2011/951403> PMID: 21766035
125. Murialdo G, Barreca A, Nobili F, Rollero A, Timossi G, Gianelli M V., et al. Relationships between cortisol, dehydroepiandrosterone sulphate and insulin-like growth factor-I system in dementia. *J Endocrinol Invest*. 2001; 24: 139–46. <https://doi.org/10.1007/BF03343833> PMID: 11314741
126. Carro E, Trejo JL, Gerber A, Loetscher H, Torrado J, Metzger F, et al. Therapeutic actions of insulin-like growth factor I on APP/PS2 mice with severe brain amyloidosis. *Neurobiol Aging*. 2006; 27: 1250–7. <https://doi.org/10.1016/j.neurobiolaging.2005.06.015> PMID: 16183170
127. Adamis D, Treloar A, Martin FC, Gregson N, Hamilton G, Macdonald AJD. APOE and cytokines as biological markers for recovery of prevalent delirium in elderly medical inpatients. *Int J Geriatr Psychiatry*. 2007; 22: 688–694. <https://doi.org/10.1002/gps.1732> PMID: 17203511
128. Preische O, Schultz SA, Apel A, Kuhle J, Kaeser SA, Barro C, et al. Serum neurofilament dynamics predicts neurodegeneration and clinical progression in presymptomatic Alzheimer's disease. *Nat Med*. 2019; 25: 277. <https://doi.org/10.1038/s41591-018-0304-3> PMID: 30664784
129. Zetterberg H, Schott JM. Biomarkers for Alzheimer's disease beyond amyloid and tau. *Nat Med*. 25 pp 201–203. 2019;25: 201–203. <https://doi.org/10.1038/s41591-019-0348-z> PMID: 30728536
130. Thompson AGB, Mead SH. Review: Fluid biomarkers in the human prion diseases. *Molecular and Cellular Neuroscience*. 2019. <https://doi.org/10.1016/j.mcn.2018.12.003> PMID: 30529227
131. Khalil M, Teunissen CE, Otto M, Piehl F, Sormani MP, Gatteringer T, et al. Neurofilaments as biomarkers in neurological disorders. *Nature Reviews Neurology*. 2018. <https://doi.org/10.1038/s41582-018-0058-z> PMID: 30171200