

1 **Impact of ozone assisted ultrasonication pre-treatment on anaerobic digestibility of**
2 **sewage sludge**

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Abbreviations: *CST*: Capillary suction time (second); *Em*: emission wavelength (nm); *Ex*: excitation wavelength (nm); *FA*: fulvic acid; *HA*: humic acid; *MW*: molecular weight (Da); *OLR*: organic loading rate (g COD/L day); *Rt*: retention time (min); *SMP*: soluble microbial products; *SRT*: solids retention time (day); *ULS*: ultrasonication; *ULS-Ozone*: ultrasonication-ozonation; *WAS*: waste activated sludge;

14 **Abstract:** Impact of ultrasonication (ULS) and ultrasonication-ozonation (ULS-Ozone)
15 pre-treatment on the anaerobic digestibility of sewage sludge was investigated with semi-
16 continuous anaerobic reactors at SRTs of 10 and 20 days. The control, ULS and ULS-
17 Ozone reactors produced 256, 309 and 348 mL biogas/g COD_{fed} and the VS removals were
18 35.6, 38.3 and 42.1%, respectively at SRT of 10 days. At SRT of 20 days, the biogas yields
19 reached 313, 337 and 393 mL/g COD_{fed} and the VS removal rates were 37.3, 40.9 and 45.3%
20 in the control, ULS and ULS-Ozone reactors, respectively. ULS-Ozone pre-treatment
21 increased the residual organics amount in the digested sludge. These soluble residual
22 organics were found to contain macromolecules with molecular weights (MW) larger than
23 500 kDa and smaller polymeric products with MW around 19.4 and 7.7 kDa. These
24 compounds were further characterized to be humic acid-like substances with fluorescent
25 spectroscopy analysis.

26 **Keywords:** Sewage sludge; ultrasound; ozone; pre-treatment; molecular weight;
27 anaerobic digestion

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31 **Introduction**

32 Ultrasonication (ULS) has been reported to be an effective sludge pre-treatment (i.e.
33 treatment of pre-digestion feed sludge) technology (Tiehm et al., 1997; Tiehm et al., 2001).
34 Biological flocs in the sludge matrix would be mechanically disrupted, resulting in particle
35 size reduction and solubilization of extra/intra- cellular polymeric substances (Bougrier et
36 al., 2005; Wang et al., 2006b). Consequently, methane production and solids removal
37 efficiency during the subsequent sludge anaerobic digestion is improved (Tiehm et al., 1997;
38 Tiehm et al., 2001). In spite of its advantages, ULS pre-treatment has limitation because it
39 is essentially “single” effect - mechanical disintegration (Khanal et al., 2007; Lehne et al.,
40 2001). Enhancement of the effectiveness of ULS pre-treatment had been attempted by
41 combining ULS process with chemical pre-treatment methods. Combination of ULS pre-
42 treatment with alkaline (Chiu et al., 1997; Jin et al., 2009; Kim et al., 2010) and acidic pre-

43 treatments (Liu et al., 2008; Sahinkaya, 2014) have been demonstrated to increase sludge
44 disintegration as well as the subsequent anaerobic digestion.

45 Apart from the aforementioned chemical methods, ozone has also been shown
46 feasible to enhance the ultrasonic pre-treatment (Tian et al., 2014; Xu et al., 2010; Yang et
47 al., 2012; Yang et al., 2013). Xu et al. (2010) demonstrated the feasibility of combining
48 ultrasound and ozone to disintegrate waste activated sludge (WAS) and to improve the
49 methane recovery from the subsequent anaerobic digestion. Yang et al. (2013) observed the
50 combined ultrasound and ozone pre-treatment enhanced the solubilization of amino acids
51 and proteins in WAS. Tian et al. (2014) found ozone was able to chemically degrade
52 macromolecules solubilized by ultrasound and further increased the sludge anaerobic
53 biodegradability.

54 These previous studies had focused on characteristics of the solubilized compounds
55 and changes in sludge properties after pre-treatment. Xu et al. (2010) and Tian et al. (2014)
56 did, however, investigate the influence of pre-treatment on sludge anaerobic
57 biodegradability in batch serum bottle tests. Information on the influence of such combined
58 pre-treatment on solids removal efficiency and digested sludge characteristics after
59 anaerobic digestion is not available. Besides, influence of solids retention time (SRT), an
60 important design parameter, on the anaerobic digestion of the combined pre-treated sludge
61 has also not been reported. This work aims to investigate the impact of such pre-treatment
62 on the subsequent anaerobic digestion process with semi-continuous reactors at SRT of 10
63 and 20 days. Biogas production and solids concentrations in the digested sludge were
64 monitored to assess the possible enhancement with such pre-treatment. Molecular weight
65 (MW) distribution and fluorescent spectroscopy analysis were conducted to provide more
66 information on the soluble residual organics in the digested sludge.

67

68 **1. Materials and methods**

69 **1.1. Sludge samples**

70 Samples of a mixture of primary sludge and thickened WAS (ratio around 1:1 based
71 on dry solids) were collected from a local municipal wastewater reclamation plant. The
72 characteristics of the sewage sludge samples are as shown in Table 1.

73

74 **1.2. Analytical methods**

75 COD and solids concentrations were measured in accordance with Standard
76 Methods (APHA, 1998). Sludge dewaterability was measured with capillary suction time
77 (CST) as described in Standard Methods (APHA, 1998). Sludge pH was measured with a
78 pH meter (Agilent, model 3200P). A UV spectrophotometer (Shimadzu, UV-1800) was
79 used in the determination of proteins, carbohydrates and ammonia-nitrogen concentrations.
80 Proteins concentration was determined with Lowry's method (1951). Carbohydrates
81 concentration was determined colorimetrically with the phenol-sulphuric acid method
82 (DuBois et al., 1956). Ammonia-nitrogen was measured colorimetrically using Nessler's
83 reagent. VFAs concentration was analysed with a gas chromatograph (Agilent
84 Technologies 7890A GC system) fitted with a flame ionization detector. The composition
85 of biogas was measured with a gas chromatograph (Agilent Technologies 7890A GC
86 system) with thermal conductivity detectors.

87

88 **1.3. Pre-treatment conditions**

89 The pre-treatment conditions were selected following consideration of the results of
90 a previous study (Tian et al., 2014). ULS pre-treatment was performed with an
91 ultrasonicator (Misonix, Q700) with ultrasound frequency of 20 kHz. The temperature was
92 monitored and maintained at about 30 °C with an ice-water bath during ultrasonication. The
93 specific energy input was 9 kJ/g TS. Ozonation pre-treatment was performed with an ozone
94 generator (WEDECO, GSO 30). Pure oxygen was used as feed gas and converted to ozone
95 with a high voltage converter. A stone diffuser was used to produce fine ozone bubbles and
96 enhance ozone mass transfer. The applied ozone dosage was 0.012 g O₃/g TS.
97 Ultrasonication-ozonation (ULS-Ozone) pre-treatment was performed by sequentially
98 applying the ULS and the ozonation treatments at the aforementioned dosages.

99

100 **1.4. Molecular weight distribution**

101 MW distribution was measured in accordance with Tian et al. (2014). A HPLC
102 (Agilent Technologies 1260 LC system) was used for MW distribution analysis using the

103 PL aquagel-OH 8 μ m MIXED-M column. Milli-Q water was used as mobile phase with a
104 flow rate of 1 mL/min. A PL aquagel-OH 8 μ m guard column was installed in front of the
105 main column. The sample was first centrifuged at 10,000 r/min for 10 min and then filtered
106 through a 0.2 μ m membrane filter before injection. A UV (254 nm) detector was used for
107 detection of the eluted substances. Corresponding MW of a detected peak was calculated by
108 converting its retention time to the corresponding MW as shown in Eq. (1) (Tian et al.,
109 2014):

$$110 \quad \text{Log}(MW) = 9.8223 - 0.6748(\text{Rt}) \quad (1)$$

111 where, Rt (min) is the retention time of the detected peaks and MW (Da) is the molecular
112 weights of the compounds detected in the corresponding peaks.

113

114 **1.5. EEM fluorescence spectroscopy analysis**

115 A fluorescence spectrometer (LS 55, Perkin Elmer, USA) was used to measure the
116 fluorescence intensity (FI) of the soluble fluorescent products. The measurement procedure
117 was previously described by Wu et al. (2011). Excitation wavelength (Ex) was from 230 to
118 520 nm with 5-nm intervals. Emission wavelength (Em) was collected from 230 to 550 nm
119 with 5-nm increments. Samples were pre-diluted 10 times with DI water to avoid the
120 measured FI exceeding the maximum level.

121 The compounds were identified based on their Ex and Em wavelengths as
122 summarized by Chen et al. (2003). Peaks of simple protein-like substances appeared in the
123 Ex/Em range of Ex < 250 nm, Em < 350 nm. Peaks of soluble microbial product (SMP)-
124 like substances were detected in the Ex/Em range of Ex: 250-280 nm, Em < 380 nm. Fulvic
125 acid (FA)-like substances fell into the Ex/Em range of Ex < 250 nm, Em > 380 nm and
126 Humic acid (HA)-like substances were detected in the Ex/Em range of Ex > 250 nm, Em >
127 380 nm.

128

129 **1.6. Anaerobic digestion tests**

130 Anaerobic digestion was conducted semi-continuously in 1.2 L glass bottles with 1
131 L working volume at 35 °C. Seed sludge was taken from a continuous anaerobic digester
132 with SRT of 28 days from a local reclamation plant. One litre of seed sludge was fed into

133 the reactor before starting the experiment. 100 and 50 mL sludge aliquots were daily
134 removed daily from the reactors and replaced with the same amount of feed sludge to
135 obtain SRT of 10 and 20 days, respectively. The reactors operating at SRT of 10 days were
136 referred to as Control10, ULS10 and ULS-Ozone10 and these received the untreated, ULS
137 treated, and ULS-Ozone treated sewage sludge as feed, respectively. Similarly, the reactors
138 at SRT of 20 days were referred to as Control20, ULS20 and ULS-Ozone20. Each reactor
139 was run for three SRTs so that process stability may be assumed. Biogas was collected with
140 Tedlar gas bags and volume measured daily with a Gas meter (Ritter, Germany). Feed
141 sludge in storage was changed every three weeks. Each batch of feed sludge was manually
142 adjusted to keep a consistent TS concentration of around 15 g/L. Daily biogas production
143 was normalised by dividing daily gas production by amount of COD fed into the reactor.

144

145 **2. Results and discussion**

146 **2.1. Biogas production and solids removal**

147 Biogas production from the anaerobic reactors is as shown in **Fig. 1a and 1b**.
148 Anaerobic biodegradability of sludge was higher at SRT of 20 days with its longer
149 substrate-microbe contact time. At both SRTs, the daily biogas production was higher from
150 the reactors fed with pre-treated sludge than from the control reactor. Average daily biogas
151 production from each reactor is as shown in **Table 2**. These values were calculated by
152 averaging the daily biogas production in the third SRT. At SRT of 10 days, daily biogas
153 production increased from 256 to 309 (+ 20.7%) and 348 (+ 35.9%) mL biogas/g COD_{fed}
154 because of the ULS and ULS-Ozone treatments of feed sludge, respectively. At SRT of 20
155 days, daily biogas production increased from 313 to 337 (+ 7.7%) and 393 (+25.5%) mL
156 biogas/g COD_{fed} due to ULS and ULS-Ozone treatments of feed sludge, respectively. These
157 results indicated that the subsequent ozonation enhanced ULS pre-treatment in terms of
158 increasing biogas production. Nickel and Neis (2007) found the improvement in biogas
159 production due to ULS treatment of feed sludge was higher when the reactor was operated
160 at a shorter SRT. For example, biogas production increased by 16% after ULS treatment of
161 feed sludge when the anaerobic reactor was operated at 8 days SRT; while the same ULS
162 treatment condition only resulted in 11% increase in biogas production when the SRT was

163 16 days. Similar results were obtained in this work. The ULS treatment of feed sludge
164 increased biogas production by 20.7% and 7.7% at SRT of 10 and 20 days, respectively. In
165 addition, results from this work showed the increase in biogas production after the ULS-
166 Ozone treatment of feed sludge also became more pronounced when the SRT was
167 shortened from 20 days to 10 days (from 25.5% to 35.9%) which had not been reported
168 previously. In all the reactors, the methane content was relatively stable at around 65%. In
169 addition, no VFAs accumulation was observed and pH value remained near neutral
170 throughout the anaerobic digestion test (around 7.0 to 7.2) in all the reactors. This
171 suggested neither ULS nor ULS-Ozone treatments of feed sludge caused stress on the
172 methanogenesis step in the reactors.

173 Improvement in organic solids removal efficiency during anaerobic digestion was
174 also observed when feed sludge was treated before anaerobic digestion. The change in VS
175 and VSS concentrations in the digested sludge during anaerobic digestion is as shown in
176 **Fig. 2a-d**. Solids concentration became relatively stable after 18 days and 30 days of
177 operation for reactors with SRT of 10 and 20 days, respectively. After reaching a relatively
178 stable level, the VS and VSS concentrations in the digested sludge were averaged for
179 comparison. The average post-digestion VS and VSS concentrations and the corresponding
180 VS and VSS removal efficiencies against the untreated feed sludge are as shown in **Table 2**.
181 The control reactor had VS removal efficiency of only 35.6% when it was operated at SRT
182 of 10 days. With the incorporation of ULS and ULS-Ozone treatments of feed sludge, the
183 VS removal rates increased to 38.3% and 42.1%, respectively. Solids removal efficiency
184 was higher at the longer SRT. The VS removal rates of the control20, ULS20 and ULS-
185 Ozone20 reactors were 37.3%, 40.9% and 45.3%, respectively. Higher VS removal
186 efficiency indicated more organic matters were digested and converted into biogas.
187 Similarly, incorporation of the pre-treatment step also improved the VSS removal
188 efficiency as shown in **Table 2**. The increase in VSS removal efficiency indicated
189 particulate organics in the treated feed sludge were better hydrolysed for the subsequent
190 anaerobic digestion process. It has been reported in a full-scale study that ULS treatment of
191 feed sludge was able to slightly decrease the VSS concentration in the digested sludge from
192 9,930 to 9,810 mg/L at SRT of 30 days (Xie et al., 2007). However, the improvement in

193 VSS removals after anaerobic digestion due to ULS treatment of feed sludge was more
194 obvious at shorter SRT of 10 and 20 days in this work (e.g. from 8,005 to 7,640 mg/L at
195 SRT of 10 days). Furthermore, this work demonstrated ULS-Ozone treatment of feed
196 sludge resulted in a lower VSS concentration in the digested sludge than ULS treatment
197 which had not been reported in previous studies.

198 T-test at the significance level of 0.05 was conducted to compare the changes in
199 biogas production and post-digestion VS concentration after the ULS and ULS-Ozone
200 treatments of feed sludge (Rivero et al., 2006; Takashima, 2008). As shown in Table 3, the
201 biogas production was significantly higher and the post-digestion VS concentration was
202 significantly lower than the control after the ULS and ULS-Ozone treatments of feed
203 sludge. In addition, the t-test results showed ULS-Ozone treatment of feed sludge resulted
204 in statistically higher biogas production and lower VS concentration in the digested sludge
205 than the ULS treatment. This confirmed application of ozonation subsequent to
206 ultrasonication could significantly enhance the sludge anaerobic digestion from a statistical
207 point of view. In addition, it was noted the daily biogas production, solids removal
208 efficiencies of the ULS-Ozone10 reactor were better than those of the Control20 reactor.
209 However, daily biogas production and solids removal rates of the ULS10 reactor were
210 lower than those of the Control20 reactor. This suggested the ULS-Ozone treatment of feed
211 sludge could halve the SRT without affecting digestion performance; whereas, the
212 individual ULS pre-treatment was not able to provide such advantage.

213

214 **2.2. Characteristics of digested sludge**

215 **2.2.1. SCOD and soluble biopolymers**

216 The SCOD concentration in the digested sludge during anaerobic digestion is shown
217 in **Fig. 3a and 3b**. At both SRTs, ULS treatment of feed sludge increased the post-
218 digestion SCOD concentration and the post-digestion SCOD increased further when ULS-
219 Ozone treatment was applied to feed sludge. The increase in SCOD in the digested sludge
220 was also compared statistically with the t-test as shown in Table 3. The statistical results
221 suggested the SCOD in the digested sludge from the ULS and ULS-Ozone reactors were
222 significantly higher than that from the control reactor. Furthermore, t-test between the

223 SCOD in the digested sludge from the ULS and ULS-Ozone reactors showed the increase
224 in post-digestion SCOD due to the subsequent ozonation was also statistically significant.
225 As shown in **Fig. 3a and 3b**, SCOD in the digested sludge from ULS-Ozone reactors were
226 around 300 and 200 mg/L higher than that from ULS reactors operating at SRT of 10 and
227 20 days, respectively; while, the SCOD in the ULS-Ozone treated feed sludge was 1,200
228 mg/L higher than the SCOD in the ULS treated feed sludge. This indicated much of the
229 COD solubilized by the subsequent ozonation treatment of feed sludge was biodegraded
230 and only a relatively small fraction accumulated in the anaerobic reactors.

231 It is known that biopolymers are a major component of sludge (Rittman and
232 McCarty, 2001). Averaged values of soluble carbohydrates and proteins concentration in
233 the digested sludge during the last three sampling days are compared in **Fig. 3c and 3d**.
234 Soluble carbohydrates and proteins concentrations in the digested sludge from the ULS-
235 Ozone reactor were much higher than the corresponding concentrations in the digested
236 sludge from the ULS and control reactor at both SRTs. This suggested undigested
237 biopolymers contributed to the higher SCOD in the digested sludge from the ULS-Ozone
238 reactors.

239 The influence of SRT on the residual carbohydrates and proteins concentrations
240 were different. As shown in **Fig. 3c**, soluble carbohydrates concentrations decreased
241 obviously when all the reactors had longer residence time. For example, the residual
242 soluble carbohydrates concentration decreased from 62 to 35 mg/L when SRT of the ULS-
243 Ozone reactor increased from 10 to 20 days. This is because the solubilized carbohydrates
244 after the treatments of feed sludge were mainly complex polysaccharides from extra- and
245 intra- cellular structures (Tian et al., 2014; Wang et al., 2006b). Longer residence time was
246 needed for sufficient degradation. However, the soluble proteins concentrations in the
247 digested sludge did not show obvious difference between SRT of 10 and 20 days for all the
248 reactors as shown in **Fig. 3d**. The residual proteins were likely to be functional proteins or
249 enzymes which could not be degraded via microbial utilization (Park et al., 2008). In
250 addition, HA which were generated during the anaerobic digestion could also be mistakenly
251 detected as proteins with the Lowry's method used.

252

253 **2.2.2. Dewaterability**

254 The ULS and ULS-Ozone treatments of feed sludge were detrimental to the
255 dewaterability of the digested sludge as shown in **Fig. 4a**. At SRT of 10 days, CST of the
256 digested sludge from the control reactor, ULS reactor and ULS-Ozone reactor were 56.9,
257 145.6 and 179.1 s, respectively. Dewaterability of the digested sludge further improved at a
258 longer residence time. At SRT of 20 days, CST of the digested sludge from the control
259 reactor, ULS reactor and ULS-Ozone reactor were 50.3, 101.4 and 120 s, respectively. This
260 was because treatment of feed sludge solubilized biopolymers which could bind with free
261 water and worsen the sludge dewaterability (Wang et al., 2006a). Some of these
262 biopolymers were persistent after anaerobic digestion and deteriorated the dewaterability of
263 the digested sludge.

264 At the same SRT, CST of the digested sludge from the ULS-Ozone reactor was
265 slightly higher than that from the ULS reactor. And, the digested sludge from the control
266 reactor had the lowest CST in comparison to the digested sludge from the ULS and ULS-
267 Ozone reactors. This indicated the dewaterability of digested sludge was deteriorated by the
268 ULS treatment of feed sludge and was further worsened by the subsequent ozonation to
269 ULS pre-treatment. Although influence of ULS-Ozone treatment of feed sludge on the
270 dewaterability of digested sludge has not been reported, results of this work were in
271 accordance with observations in a previous study where individual ULS and individual
272 ozone treatments of feed sludge were found to deteriorate the dewaterability of digested
273 sludge (Braguglia et al., 2012).

274 **2.2.3. Ammonia-nitrogen**

275 Ammonia-nitrogen concentration increased after anaerobic digestion as a result of
276 the degradation of proteinous compounds and absence of nitrogen removal pathways (Kim
277 et al., 2010). Averaged ammonia-nitrogen concentration in the digested sludge in the last
278 three days of the anaerobic digestion tests were compared in **Fig. 4b**. Digested sludge from
279 the reactors fed with treated feed sludge had higher ammonia-nitrogen concentration than
280 that from the control reactors. Previous studies indicated the increase of ammonium
281 concentration in the digested sludge could be a drawback of the pre-treatment step because
282 pre-treatment steps released intra- and extra- cellular proteins to be anaerobically degraded

283 (Dogan and Sanin, 2009; Kim et al., 2010). However, it was noted that ULS-Ozone
284 treatment of feed sludge did not have such effect on ammonia in the digested sludge of the
285 anaerobic digester compared to ULS treatment. This might be due to the oxidative effect of
286 ozone.

287

288 **2.3. Molecular weight distribution**

289 MW distribution chromatograms of the standard polymers are shown in **Fig. 5a**.
290 MW chromatograms of the soluble substances in the feed sludge, in the digested sludge
291 from reactors operating at 10 days SRT and in the digested sludge from reactors operating
292 at 20 days SRT are shown in **Fig. 5b, 5c and 5d**, respectively. Detected peaks were divided
293 into five groups (A to F) in ascending order of retention time. MWs of the components in
294 these peaks are in the descending order from A to F because larger compounds were
295 retained for a shorter time in the column and eluted earlier.

296

297 **2.3.1. High MW compounds**

298 Peak A (Rt: 4.0 min) and B (Rt: 5.6 min) had the most obvious increase after
299 treatments of feed sludge as shown in **Fig. 5b**. Compounds detected in these peaks were
300 macromolecules with MW higher than 500 kDa because retention time of these peaks was
301 shorter than the retention time of the largest tested standard polymer (MW: 500 kDa, Rt:
302 6.2 min). Therefore, these compounds were likely to be high MW extra- and intra-
303 polymeric substances released from sludge matrix after the treatments of feed sludge. MW
304 distribution of the digested sludge was shown in **Fig. 5c and 5d**. Peak C (Rt: 6.0 min, MW >
305 500 kDa) instead of peak B was detected in the digested sludge together with peak A.
306 Compounds detected in peak C could be generated from hydrolysis of particulate polymers
307 and higher MW macromolecules (peak A) by hydrolytic bacteria because peak C was
308 detected only after anaerobic digestion. It should be noted that peak C was broader than
309 peak B and covered the retention time of peak B by comparing **Fig. 5c, 5d to 5b**. Therefore,
310 peak B was possibly over-dominated by peak C and thus not detected. As a result, soluble
311 biopolymers released by treatments of feed sludge could also be detected in peak C if
312 remaining undigested.

313 At SRT of 10 days, responses of peak A and C in the digested sludge from the ULS-
314 Ozone reactor were significantly higher than the corresponding responses in the digested
315 sludge from the control and ULS reactors as shown in **Fig. 5c**. This was due to the
316 subsequent ozonation process because such response increase was not observed when only
317 ULS treatment was applied to feed sludge. Similar observations were made on peak D (Rt:
318 8.2 min) and peak E (Rt: 8.8 min) with MW around 19.4 kDa and 7.7 kDa, respectively.
319 These compounds (detected in peak D and E) were most likely to be intermediate products
320 generated during the anaerobic degradation of macromolecules into monomers because
321 their amounts were significantly lower in the feed sludge than those in the digested sludge.

322 Responses of peak A, C, D and E were lower at SRT of 20 days as compared in **Fig.**
323 **5c and 5d**. This indicated some of the compounds detected in these peaks were slowly
324 biodegradable compounds. They were not biodegraded at SRT of 10 days but could be
325 digested at SRT of 20 days. Some of these compounds were likely to be carbohydrates
326 because the chemical results determined in Section 2.2.1 showed some carbohydrates were
327 complex polysaccharides and were not biodegradable at SRT of 10 days but became
328 biodegradable at SRT of 20 days. At SRT of 20 days, no obvious difference was observed
329 between the MW chromatograms of the digested sludge from the control and ULS reactors
330 as shown in **Fig. 5d**. However, responses in peak A, C, D and E were significantly higher
331 in the digested sludge from the ULS-Ozone reactor. This indicated considerable amounts of
332 polymeric substances remained undigested in the ULS-Ozone reactor even at SRT of 20
333 days. These residual soluble polymeric compounds in the anaerobic digested sludge were
334 possibly related to solubilization of persistent compounds after the ULS-Ozone treatment of
335 feed sludge (Tian et al., 2014; Yang et al., 2013). These results correlated very well with
336 the increase in biopolymers in the digested sludge as observed in Section 2.2.1.

337

338 **2.3.2. Low MW compounds**

339 Peak F (Rt: 13.3 min, < 106 Da) was detected in the supernatant of digested sludge
340 as shown in **Fig. 5c and 5d**. The corresponding compounds were not monomers or other
341 easily biodegradable components because they remained undigested at SRT of 20 days.
342 Therefore, they were possibly short chain alkenes or aromatics which are anaerobic

343 digestion by-products and could be detected by the UV 254 nm detector. Formation of
344 these by-products is related to the chemical effects of the subsequent ozonation process
345 because the response of peak F was obviously higher in the digested sludge from the ULS-
346 Ozone reactors. It is possible that the complex polymers in the untreated and ULS pre-
347 treated sludge could only be broken down via biodegradation; while, dosing ozone could
348 provide different degradation pathways by chemically breaking down the high MW
349 biopolymers into smaller fragments (Tian et al., 2014). Besides, ozone can convert some
350 refractory compounds into biodegradable ones (Nishijima et al., 2003). These
351 aforementioned factors could generate different substrates and anaerobic digestion of these
352 new substrates could contribute to the accumulation of the detected by-products.

353 Previous studies had focused on VSS removal and biogas production increase due to
354 treatment of feed sludge, but insights on the SCOD in the anaerobic digested sludge were
355 not discussed in these studies (Dogan and Sanin, 2009; Tiehm et al., 2001). SCOD in the
356 digested sludge could be attributed to slowly degradable components and recalcitrant
357 anaerobic digestion by-products. MW distribution results allow better realization of the
358 possible sources and categories of the residual components according to their MWs. These
359 would be good supplementary information to conventional approaches in understanding the
360 influence of treatments of feed sludge on subsequent anaerobic digestion.

361

362 **2.4. Fluorescent products characterization**

363 EEM fluorescence spectroscopy analysis was conducted to measure the
364 fluorescence intensity (FI) of fluorescent compounds in the supernatant of digested sludge
365 the reactors. The EEM spectra of all the samples are shown in **Fig. 6a-f**. FI of the detected
366 peaks was shown in numbered and coloured contour lines for reference. According to the
367 Ex/Em range introduced in Section 1.5, the main peaks were HA-like (as highlighted with a
368 white arrow in **Fig. 6c**) and FA-like substances (as highlighted with a white arrow in **Fig.**
369 **6d**). These substances were released from the biodegradation of the extracellular polymeric
370 substances, sludge pellets and refractory components (e.g. lignin) in the sewage sludge
371 (Luo et al., 2013). Aside from these two groups, SMP-like substances and simple protein-

372 like matters were also detected in each spectrum. However, their FIs were relatively low
373 and over-dominated by peaks of HA-like and FA-like substances.

374

375 **2.4.1. Humic acid-like substances**

376 At SRT of 10 days, the FI of HA-like substances were similar in the digested sludge
377 from the control and ULS reactors as shown in **Fig. 6a and 6b**. In contrast, the FI of the
378 HA-like substances were significantly higher in the digested sludge from the ULS-Ozone
379 reactor as shown in **Fig. 6c**. This confirmed some proteins detected with the Lowry's
380 method in Section 2.2.1 were attributed to humic substances. This is because the ULS-
381 Ozone treatments of feed sludge disintegrated the sludge better and solubilized more HA-
382 containing substances in comparison to the ULS treatment. Biodegradation of these
383 solubilized HA-containing substances resulted in a higher concentration of HAs as by-
384 products. These HAs should contribute to the residual polymeric substances in the digested
385 sludge from the ULS-Ozone reactors (e.g Peak C, in **Fig. 5d**) as discussed in Section 2.3.1,
386 because HAs are known to be persistent and have high MW (Li et al., 2009; Stevenson,
387 1994). Such increase in HAs during anaerobic digestion of pre-treated sludge was in good
388 agreement with results obtained by Luo et al. (2013). They observed anaerobic digestion of
389 enzymatically pre-treated WAS resulted in higher FI of HA-like substances in the digested
390 sludge compared to anaerobic digestion of untreated WAS (Luo et al., 2013).

391 By comparing the EEM spectra in **Fig. 6a, 6b** to **Fig. 6d, 6e**, FI of the HA-like
392 matters in the digested sludge from the control and the ULS reactors were both found to
393 increase at the longer retention time. This was likely because sludge were better digested at
394 the longer retention time which released more HAs as anaerobic digestion by-products. In
395 contrast, FI of the HA-like substances were similar in the digested sludge of the ULS-
396 Ozone10 and the ULS-Ozone20 reactors as shown in **Fig. 6c and 6f**. This indicated the
397 HA-containing substances were mostly biodegraded and the HAs were released into the
398 supernatant within 10 days of anaerobic digestion for the ULS-Ozone pre-treated sludge. A
399 longer digestion time did not further increase the FI of the HA-like substances in the
400 anaerobic digested sludge.

401

402 **2.4.2. Fulvic acid-like substances**

403 FI of the FA-like substances were similar in the digested sludge from the control,
404 ULS and ULS-Ozone reactors at SRT of 10 days as shown in **Fig. 6a, 6b and 6c**. By
405 comparing **Fig. 6a, 6b** to **Fig. 6d, 6e**, FI of the FA-like substances in the digested sludge of
406 the control and the ULS reactors were both found to increase when the SRT increased to 20
407 days which was similar to the observations on the HA-like matters. However, the FI of the
408 FA-like substances in the digested sludge of the ULS-Ozone reactor decreased when the
409 SRT increased from 10 days to 20 days, indicating the FA-like compounds became
410 biodegradable at a longer digestion time due to the subsequent ozonation step. This was
411 supported by previous studies which observed the increase of biodegradability of FAs due
412 to ozonation process (Kozyatnyk et al., 2013; Volk et al., 1997). Such increase in
413 biodegradability of FAs is a potential advantage of ozonation treatment of feed sludge and
414 has not been emphasized in previous studies.

415

416 **3. Conclusions**

417 This work investigated the impact of ULS-Ozone treatment of pre-digestion feed
418 sludge on sludge anaerobic digestion. The findings of this work are summarized as follows:

- 419 • Subsequent ozonation complemented ULS treatment in improving biogas
420 production and volatile solids removal when the feed sludge was treated
421 before anaerobic digestion.
- 422 • ULS-Ozone treatment of feed sludge could shorten the anaerobic digestion
423 SRT from 20 days to 10 days without adverse impact on anaerobic digestion
424 performance.
- 425 • Soluble polymeric substances were found to accumulate in the anaerobic
426 digested sludge following anaerobic digestion of ULS-Ozone treated feed
427 sludge. Such digested sludge had deteriorated dewaterability. Although
428 some of these polymers were anaerobically degradable at 20 days SRT, most
429 were HA-like substances and persistent.
- 430 • Biodegradability of FA-like substances was improved due to application of
431 ozone.

432

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435 Singapore for sponsoring the project and providing sludge samples for analysis and Xylem
436 Water Solutions Herford GmbH for providing the ozone generator.

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538 waste activated sludge in dissolved organic matter following ultrasound
539 assisted ozone pretreatments. *Bioresour. Technol.*, 131(0), 560-563.
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542

543 **List of Tables:**

544 **Table 1 Main characteristics of the untreated sewage sludge**

Parameters	Value range
Total solids (g/L)	14.9 - 15.4
Total suspended solids (g/L)	13.7 – 14.1
Volatile solids (g/L)	12.7 - 13.2
Volatile suspended solids (g/L)	11.8 – 12.0
Total COD (g/L)	17.4 – 20.0
Soluble COD (g/L)	0.82 – 1.26
pH	5.9 – 6.2

545

546

Table 2 Performance of the semi-continuous anaerobic digesters at the assumed steady state

Reactor	SRT 10 days			SRT 20 days		
	Control10	ULS10	ULS-Ozone10	Control20	ULS20	ULS-Ozone20
Organic loading rate (g COD/L.day) (n=10 and 20) ^a	1.78 ± 0.05	1.71 ± 0.04	1.62 ± 0.02	0.99 ± 0.02	0.97 ± 0.01	0.93 ± 0.02
Biogas (mL/day g COD _{fed}) (n=10 and 20) ^a	256 ± 5	309 ± 6	348 ± 9	313 ± 7	337 ± 10	393 ± 12
Methane percentage (%) (n=5)	64.2 ± 0.9	65.2 ± 0.8	65.6 ± 0.6	64.9 ± 0.5	64.9 ± 1.3	65.4 ± 1.2
VS in digested sludge (mg/L) (n=5 and 8) ^b	8340 ± 383	7990 ± 344	7500 ± 245	8125 ± 252	7644 ± 253	7081 ± 217
VS removal efficiency (%) (n=5 and 8) ^b	35.6 ± 4.6	38.3 ± 4.3	42.1 ± 3.3	37.3 ± 3.1	40.9 ± 3.3	45.3 ± 3.1
VSS in digested sludge (mg/L) (n=5 and 8) ^b	8005 ± 469	7640 ± 155	6760 ± 248	7619 ± 258	7056 ± 213	6606 ± 206
VSS removal efficiency (%) (n=5 and 8) ^b	32.7 ± 3.9	35.8 ± 1.3	43.2 ± 2.1	36.0 ± 2.2	40.7 ± 1.8	44.5 ± 1.7
Total VFA in digested sludge (mg/L) (n=3)	Nd ^c	Nd	Nd	Nd	Nd	Nd

547

^a n =10 at SRT of 10 days and n= 20 at SRT of 20 days

548

^b n =5 at SRT of 10 days and n= 8 at SRT of 20 days

549

^c not detectable (< 10 mg/L)

550

551

552 **Table 3 Statistical analysis of the biogas production, VS concentration in digested sludge and SCOD in**
 553 **digested sludge after ULS and ULS-Ozone pre-treatments at different SRTs**

	ULS	ULS-Ozone
Biogas (mL/day g COD _{fed})		
Compared to control at SRT of 10 days (n=10)	t: 23.398, P: 2.27×10^{-9} ^a	t: 38.203, P: 2.86×10^{-11} ^a
Compared to ULS at SRT of 10 days (n=10)	-	t: 16.329, P: 5.39×10^{-8} ^a
Compared to control at SRT of 20 days (n=20)	t: 12.709, P: 9.77×10^{-11} ^a	t: 33.657, P: 2.11×10^{-18} ^a
Compared to ULS at SRT of 20 days (n=20)	-	t: 25.715, P: 3.16×10^{-16} ^a
VS concentration in digested sludge (mg/L)		
Compared to control at SRT of 10 days (n=5)	t: -4.111, P: 0.01473 ^b	t: -5.126, P: 0.0686 ^b
Compared to ULS at SRT of 10 days (n=5)	-	t: -3.642, P: 0.02192 ^b
Compared to control at SRT of 20 days (n=8)	t: -7.434, P: 1.45×10^{-3} ^b	t: -12.166, P: 5.80×10^{-6} ^b
Compared to ULS at SRT of 20 days (n=8)	-	t: -7.515, P: 1.36×10^{-4} ^b
SCOD in digested sludge (mg/L)		
Compared to control at SRT of 10 days (n=5)	t: 8.573, P: 0.00102 ^a	t: 24.305, P: 1.70×10^{-5} ^a
Compared to ULS at SRT of 10 days (n=5)	-	t: 24.331, P: 1.69×10^{-5} ^a
Compared to control at SRT of 20 days (n=8)	t: 4.694, P: 0.00222 ^a	t: 21.649, P: 1.13×10^{-7} ^a
Compared to ULS at SRT of 20 days (n=8)	-	t: 18.243, P: 3.68×10^{-7}

554 t is the *t statistic* value and P is the probability that the two compared values are not significantly different

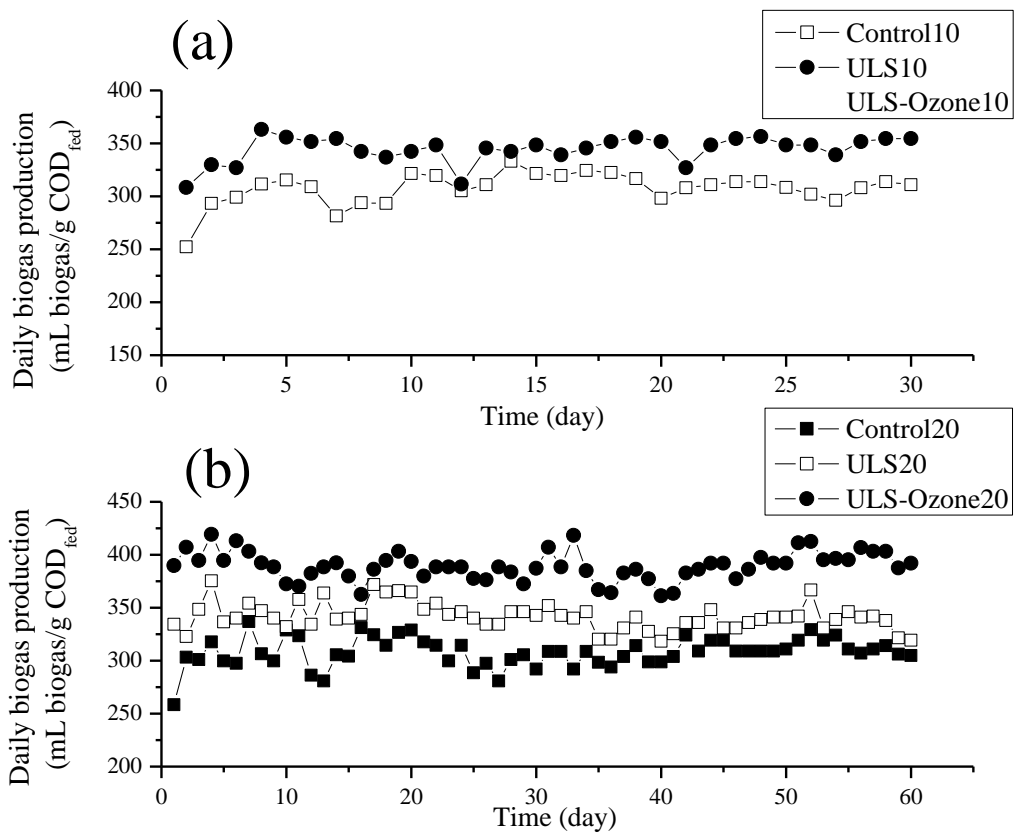
555 ^a The tested value was significantly higher than the reference value

556 ^b The tested value was significantly lower than the reference value

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558 List of figures:

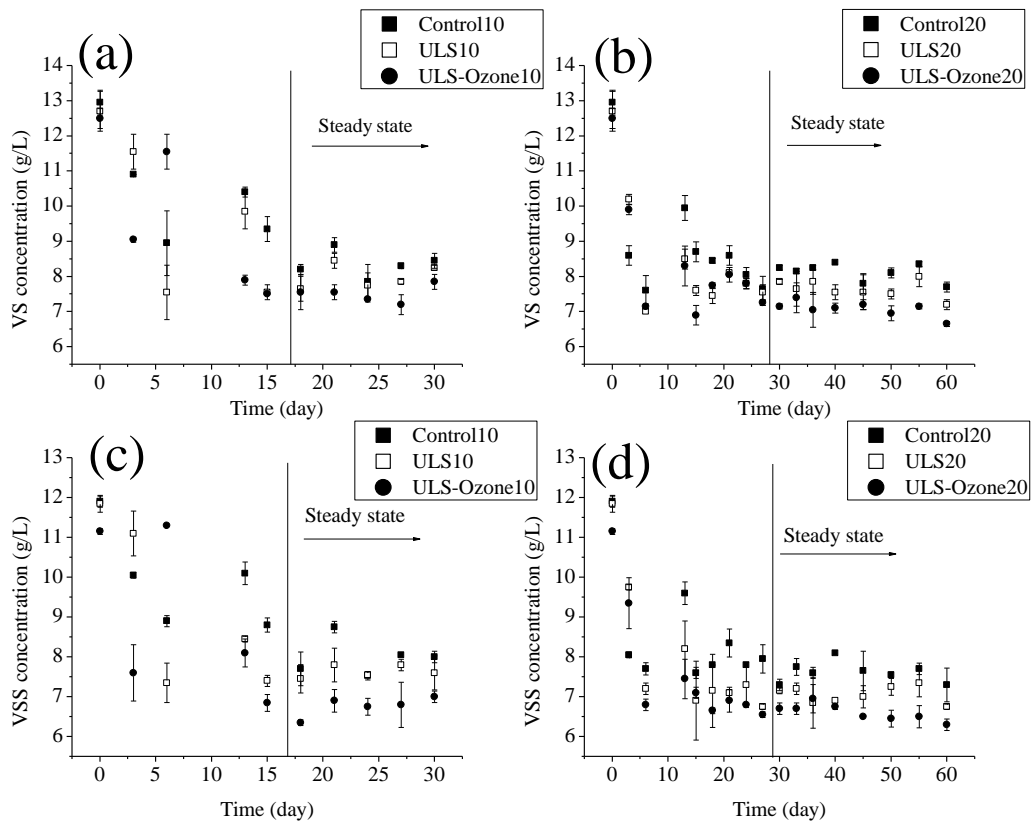
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561 Fig.1 Daily biogas production from the reactors at (a) SRT of 10 days (Control 10, ULS10, ULS-
562 Ozone10) (b) SRT of 20 days (Control 20, ULS20, ULS-Ozone20)

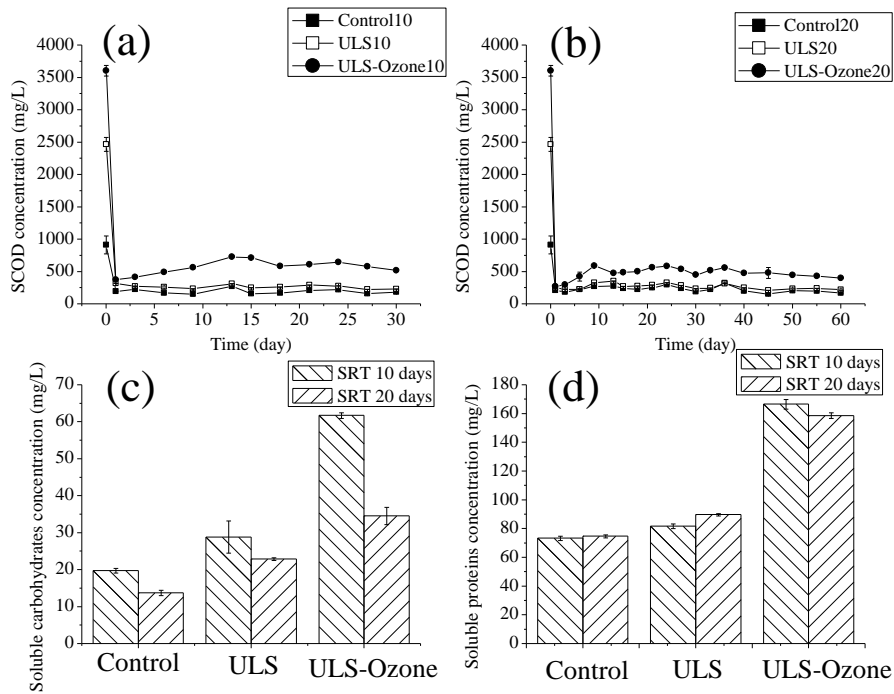
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566 **Fig. 2 (a) Change in VS concentration in the digested sludge during the anaerobic digestion at SRT of**
 567 **10 days (b) Change in VS concentration in the digested sludge during the anaerobic digestion at SRT of**
 568 **20 days (c) Change in VSS concentration in the digested sludge during the anaerobic digestion at SRT**
 569 **of 10 days (d) Change in VSS concentration in the digested sludge during the anaerobic digestion at**
 570 **SRT of 20 days (day 0 stands for feed sludge) (solid concentration was based on at least two replicates)**

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572

573 **Fig. 3 (a) Change in SCOD concentration in the digested sludge during the anaerobic digestion at SRT**

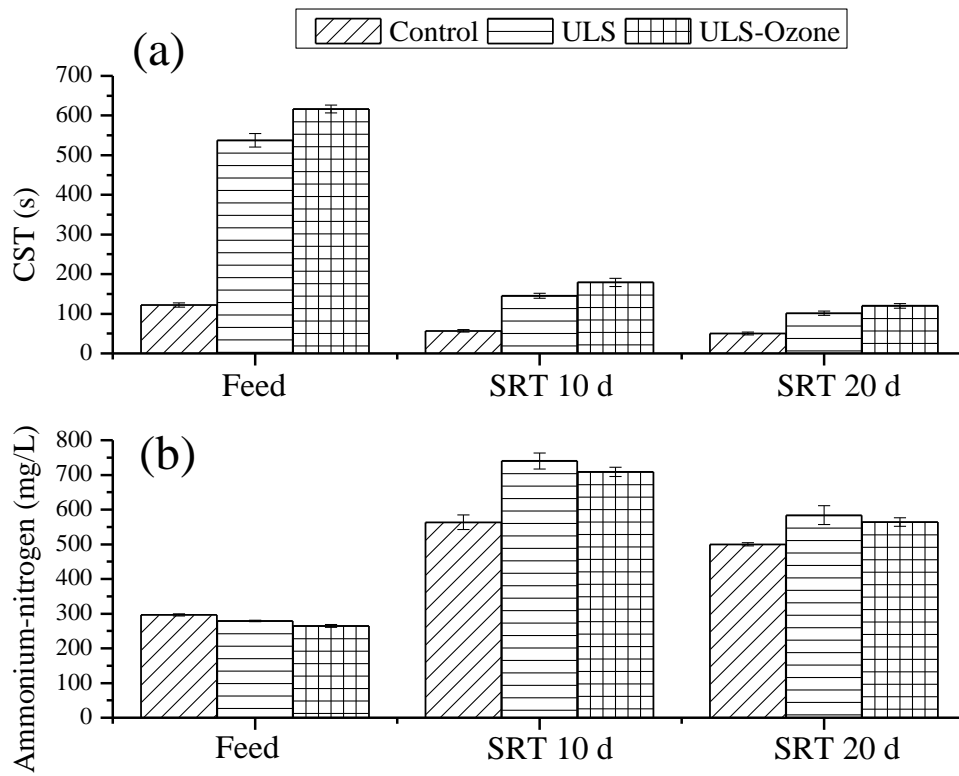
574 **of 10 days (b) Change in SCOD concentration in the digested sludge during the anaerobic digestion at**

575 **SRT of 20 days (SCOD concentration was based on at least two replicates) (c) Soluble carbohydrates**

576 **concentrations in the digested sludge from different anaerobic reactors (n=3) (d) Soluble proteins**

577 **concentrations in the digested sludge from different anaerobic reactors (n=3)**

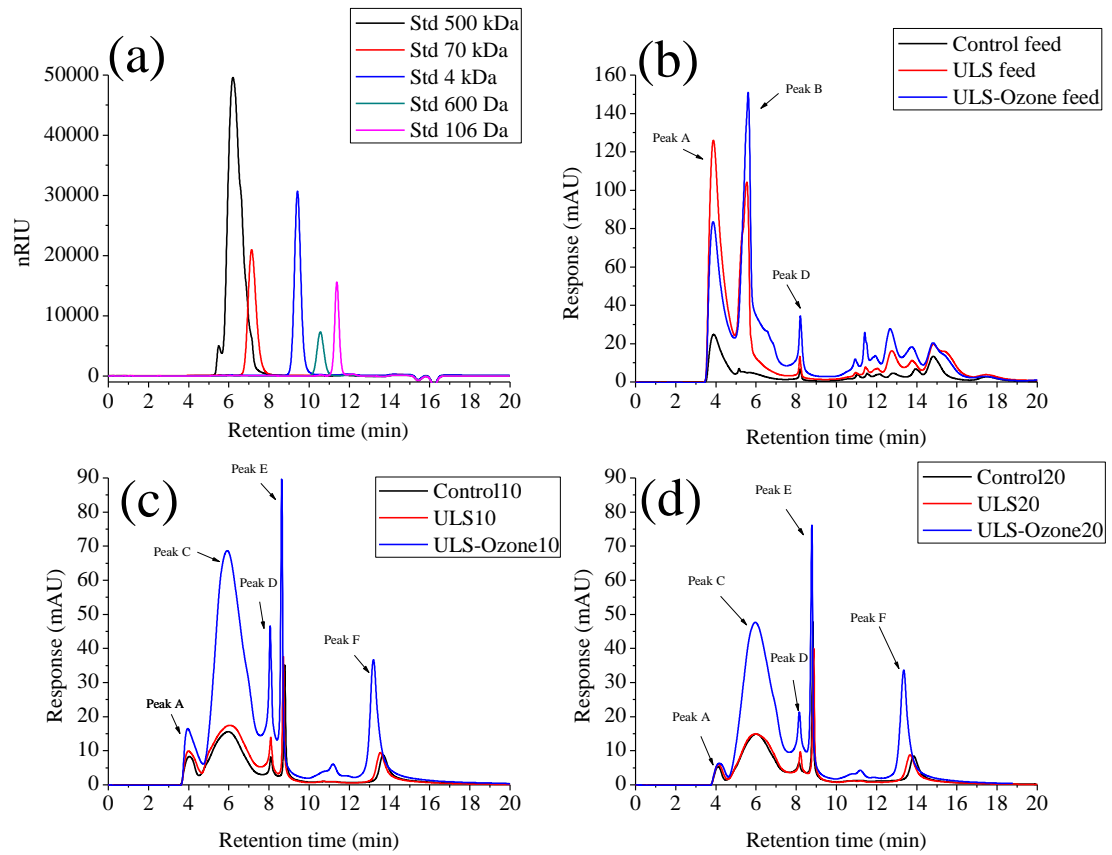
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580 **Fig. 4 (a) Dewaterability (n=3) (b) Ammonia-nitrogen (n=3) in feed sludge and digested sludge from**
 581 **different anaerobic reactors**

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585 **Fig. 5 Molecular weight distribution chromatograms of (a) standard polymers (b) supernatant in the**

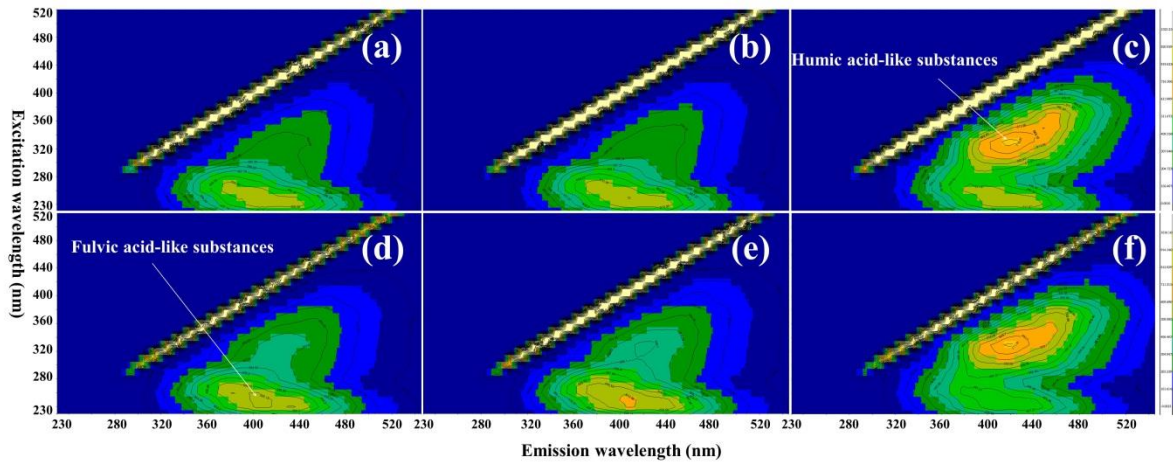
586 **feed sludge (c) supernatant in the digested sludge from reactors operating at SRT of 10 days (d)**

587 **supernatant in the digested sludge from reactors operating at SRT of 20 days**

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592 **Fig.6 Excitation emission matrix fluorescent spectra of the supernatant in the digested sludge from (a)**

593 **control10 (b) ULS10 (c) ULS-Ozone10 (d) control20 (e) ULS20 (f) ULS-Ozone20**